

Mouse MPO Immunoassay

Catalog Number: EA800166

For the quantitative determination of mouse MPO concentrations in cell culture supernates, serum, and plasma.

For research use only. Not for use in diagnostic procedures.

MANUFACTURED AND DISTRIBUTED BY:

OriGene Technologies,Inc.

9620 Medical Center Drive Suite 200 Rockville, MD 20850,USA

Tel: 1-301-340-3188 Fax: 301-340-9254 Mail: techsupport@origene.com Web: www.origene.com



TABLE OF CONTENTS

SECTION	PAGE	
BACKGROUND		.1
PRINCIPLE OF THE ASSAY		1
TECHNICAL HINTS AND LIMITATIONS		2
PRECAUTIONS	2	2
KIT COMPONENTS& STORAGE CONDITIONS		3
OTHER SUPPLIES REQUIRED BUT NOT SUPP	LIED	.4
SPECIMEN COLLECTION & STORAGE		,4
REAGENTS PREPARATION	2	4
ASSAY PROCEDURE		.6
CALCULATION OF RESULTS		6
PERFORMANCE CHARACTERISTICS		8
REFERENCES	1	0



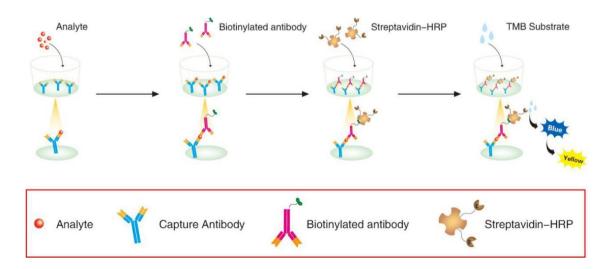
BACKGROUND

Myeloperoxidase (MPO) is a heme-containing enzyme belonging to the XPO subfamily of peroxidases. It is an abundant neutrophil and monocyte glycoprotein that catalyzes the hydrogen peroxide dependent formation of hypochlorus acid (HOCl) and other reactive species. Enzymatically active MPO is a disulfide-linked tetramer that contains two heme groups and two copies each of the heavy and light chains. MPO binds Albumin, MMR, Cytokeratin 1 on vascular endothelial cells, HMW Kiningen, and Integrin CD11b/CD18 on neutrophils. These interactions promote MPO clearance, a reduction of nitric oxide and bradykinin levels, reduced vasodilation, and continued neutrophil activation. Neutrophil MPO is stored in cytoplasmic azurophilic granules. Upon cellular activation and degranulation, MPO is delivered into phagosomes where it is required for the killing of phagocytosed bacteria. Activated neutrophils also release granule contents extracellularly. Elevated plasma MPO levels have been associated with a variety of clinical conditions including systemic inflammation, eclampsia, risk of cardiovascular events, vascular endothelial dysfunction, severity of multiple sclerosis, and prospective mortality and oxidative stress during hemodialysis.

PRINCIPLE OF THE ASSAY

This assay employs the quantitative sandwich enzyme immunoassay technique. A monoclonal antibody specific for MPO has been pre-coated onto a microplate. Standards and samples are pipetted into the wells and any MPO present is captured by the coated antibody after incubation. Following extensive washing, a biotin-conjugate antibody specific for MPO is added to detect the captured MPO protein in sample. For signal development, horseradish peroxidase (HRP)-conjugated Streptavidin is added, followed by tetramethyl-benzidine (TMB) reagent. Following a wash to remove any unbound combination, and enzyme conjugate is added to the wells. Solution containing sulfuric acid is used to stop color development and the color intensity which is proportional to the quantity of bound protein is measurable at 450nm.

Schematic diagram:





TECHNICAL HINTS AND LIMITATIONS

- 1. This ELISA should not be used beyond the expiration data on the kit label.
- 2. To avoid cross-contamination, use a fresh reagent reservoir and pipette tips for each step.
- 3. To ensure accurate results, some details, such as technique, plasticware and water sources should be emphasized.
- 4. A thorough and consistent wash technique is essential for proper assay performance.
- 5. A standard curve should be generated for each set of samples assayed.
- 6. It is recommended that all standards and samples be assayed in duplicate.
- 7. Avoid microbial contamination of reagents and buffers. Buffers containing protein should be made under aseptic conditions and be prepared fresh daily.
- 8. In order to ensure the accuracy of the results, the standard curve should be made every time.

PRECAUTIONS

The Stop Solution suggested for use with this kit is an acid solution. Wear protective gloves, clothing, eye, and face protection. Wash hands thoroughly after handling.



PART	SIZE	STORAGE OF OPENED/ RECONSTITUTED MATERIAL			
Microwell Plate - antibody coated 96-well Microplate (8 wells ×12 strips)	1 plate	Return unused wells to the foil pouch containing the desiccant pack. Reseal along entire edge of the zip-seal. May be stored for up to 1 month at $2-8^{\circ}C^{**}$			
Standard -lyophilized,8000 pg/vial upon reconstitution	2 vials	Aliquot and Store at -20°C** for six months			
Concentrated Biotin-Conjugated antibody(100X) - 120 ul/vial	1 vial	Store at 2-8°C **for six months			
Concentrated Streptavidin-HRP solution(100X) - 120 ul/vial	1 vial	Store at 2-8°C** for six months			
Standard /Sample Diluent - 16 ml/vial	1 bottle	Store at 2-8°C** for six months			
Biotin-Conjugate antibody Diluent - 16 ml/vial	1 bottle	Store at 2-8°C** for six months			
Streptavidin-HRP Diluent - 16 ml/vial	1 bottle	Store at 2-8°C** for six months			
Wash Buffer Concentrate (20x) - 30 ml/vial	1 bottle	Store at 2-8°C** for six months			
Substrate Solution - 12 ml/vial	1 bottle	Store at 2-8°C** for six months			
Stop Solution - 12 ml/vial	1 bottle	Store at 2-8°C** for six months			
Plate Cover Seals	4 pieces				

^{**}Provided this is within the expiration date of the kit.

OTHER SUPPLIES REQUIRED BUT NOT SUPPLIED

1. Microplate reader capable of measuring absorbance at 450 nm.



- 2. Pipettes and pipette tips.
- 3. Deionized or distilled water.
- 4. Squirt bottle, manifold dispenser, or automated microplate washer.
- 5. 500 mL graduated cylinder.

SPECIMEN COLLECTION & STORAGE

Cell Culture Supernates - Centrifuge cell culture media at $1000 \times g$ to remove debris. Assay immediately or aliquot and store samples at \leq -20 °C. Avoid repeated freezethaw cycles.

Serum - Use a serum separator tube (SST) and allow samples to clot for 2 hours at room temperature or overnight at 2-8°C. Centrifuge approximately for 15 minutes at $1000 \times g$. Assay immediately or aliquot and store samples at \le -20 °C. Avoid repeated freeze-thaw cycles.

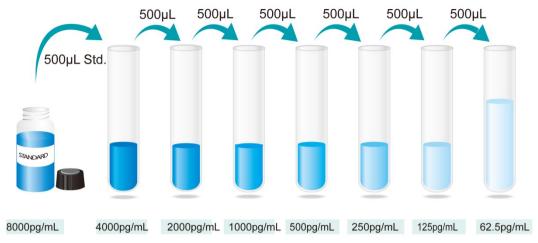
Plasma - Collect plasma using EDTA, heparin, or citrate as an anticoagulant. Centrifuge for 15 minutes at $1000 \times g$ within 30 minutes of collection. Assay immediately or aliquot and store samples at \leq -20 °C. Avoid repeated freeze-thaw cycles.

Note: The normal mouse serum or plasma samples are suggested to make a 1:2 dilution.

REAGENTS PREPARATION

- **1. Temperature returning** Bring all kit components and specimen to room temperature (20-25°C) before use.
- **2. Wash Buffer** Dilute 30mL of Wash Buffer Concentrate with 570mL of deionized or distilled water to prepare 600mL of Wash Buffer. If crystals have formed in the concentrate Wash Buffer, warm to room temperature and mix gently until the crystals have completely dissolved.
- 3. Standard/Specimen Reconstitute the Standard with 1.0mL of Standard/Sample Diluent. This reconstitution produces a stock solution of 8000 pg/mL. Allow the standard to sit for a minimum of 15 minutes with gentle agitation prior to making dilutions. Pipette 500μL of Standard/ Sample Diluent into 4000pg/ml tube and the remaining tubes. Use the stock solution of 8000pg/mL to produce a 2-fold dilution series (below). Mix each tube thoroughly and change pipette tips between each transfer. The 4000 pg/mL standard serves as the high standard. The Standard/ Sample Diluent serves as the zero standard (0 pg/mL).





Preparation of MPO standard dilutions

- *If you do not run out of re-melting standard, store it at -20°C. Diluted standard shall not be reused.
- **4. Working solution of Biotin-Conjugate anti-mouse MPO antibody:** Make a 1:100 dilution of the concentrated Biotin-Conjugate solution with the Biotin-Conjugate antibody Diluent in a clean plastic tube.
 - *The working solution should be used within one day after dilution.
- **5. Working solution of Streptavidin-HRP**: Make a 1:100 dilution of the concentrated Streptavidin-HRP solution with the Streptavidin-HRP Diluent in a clean plastic tube.
 - *The working solution should be used within one day after dilution.

ASSAY PROCEDURE

Prepare all reagents and standards as directed. Wash the plate 3 times before assay.



Add 100µl standard or samples to each well, incubate 90 minutes,37°C.



Aspirate and wash 4 times

Add 100µl working solution of Biotin-Conjugate anti-mouse MPO antibody to each well, incubate 60 minutes, 37°C.

 \square Aspirate and wash 4 times

Add 100µl working solution of Streptavidin-HRP to each well, incubate 30 minutes,37°C.

 \bigcirc Aspirate and wash 5 times

Add 100µl Substrate solution to each well, incubate 15 minutes,37°C.Protect from light.

Ţ

Add 50µl Stop solution to each well. Read at 450nm within 5 minutes.

CALCULATION OF RESULTS

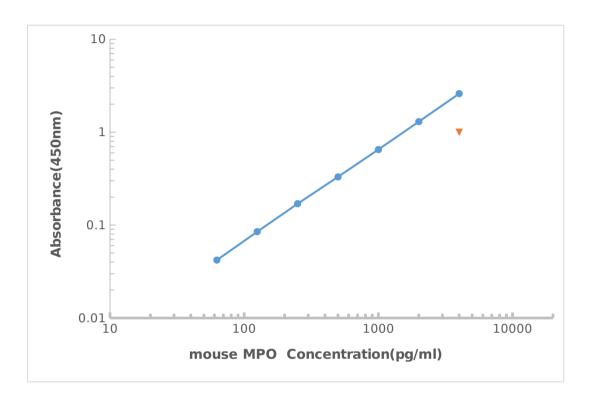
- 1. The standard curve is used to determine the amount of specimens.
- First, average the duplicate readings for each standard, control, and sample. All
 O.D. values are subtracted by the mean value of blank control before result
 interpretation.
- 3. Construct a standard curve by reducing the data using computer software capable of generating a four parameter logistic (4-PL) curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph.
- 4. The data may be linearized by plotting the log of the MPO concentrations versus the log of the O.D. and the best fit line can be determined by regression analysis. This procedure will produce an adequate but less precise fit of the data. If samples have been diluted, the concentration read from the standard curve must be multiplied by the dilution factor.
- 5. This standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.

Typical data using the MPO ELISA

Standard(pg/ ml)	OD.	OD.	Average	Corrected
0	0.084	0.073	0.078	
62.5	0.116	0.099	0.107	0.029
125	0.172	0.137	0.154	0.076



250	0.277	0.245	0.261	0.182
500	0.547	0.455	0.501	0.422
1000	0.972	0.994	0.983	0.904
2000	1.725	1.670	1.697	1.619
4000	2.336	2.301	2.318	2.240



Representative standard curve for MPO ELISA.

Performance Characteristics

SENSITIVITY: The minimum detectable dose was 30pg/mL.

SPECIFICITY: This assay recognizes both natural and recombinant mouse MPO. The factors listed below were prepared at 100ng/ml in Standard /sample Diluent and



assayed for cross-reactivity and no significant cross-reactivity or interference was observed.

Factors assayed for cross-reactivity

Recombinant mouse	Recombinant rat	Recombinant human
GPX-1		MPO
MMR		
PROS-1		

REPEATABILITY: The coefficient of variation of both intra-assay and inter-assay were less than 10%.

RECOVERY: The recovery of MPO spiked to three different levels in four samples throughout the range of the assay in various matrices was evaluated.

Recovery of MOP in two matrices

Sample Type	Average % of Expected Range (%)	Range (%)
Citrate plasma	92	86-101
Cell culture supernatants	95	84-104

LINEARITY: To assess the linearity of the assay, three samples were spiked with high concentrations of MPO in various matrices and diluted with the appropriate Sample Diluent to produce samples with values within the dynamic range of the assay. (The plasma samples were initially diluted 1:1)

Dilution ratio	Recove	ery (%)	Citrate p	lasma	Cell	culture
Tel: 1-301-3	40-3188	Mail: techsupport@oi	rigene.com	Web: w	ww.origene	.com



			supernatants
1:2	Average% of Expected	95	103
1.2	Range (%)	86-105	85-112
1.4	Average% of Expected	93	105
1:4	Range (%)	84-102	98-114
1:8	Average% of Expected	96	106
1.0	Range (%)	87-107	95-118
1.10	Average% of Expected	92	104
1:16	Range (%)	85-101	95-112

REFERENCES

- 1. Klebanoff, S.J. (2005) J. Leukoc. Biol. 77:598.
- 2. Morishita, K. et al. (1987) J. Biol. Chem. 262:3844.
- 3. van Dalen, C.J. et al. (2000) J. Biol. Chem. 275:11638.
- 4. Podrez, E.A. et al. (1999) J. Clin. Invest. 103:1547.



- 5. Bergt, C. et al. (2004) Proc. Natl. Acad. Sci. 101:13032.
- 6. Hazen, S.L. et al. (1996) J. Biol. Chem. 271:23080.
- 7. Heinecke, J.W. et al. (1993) J. Clin. Invest. 91:2866.
- 8. Kulcharyk, P.A. and J.W. Heinecke (2001) Biochemistry 40:3648.
- 9. Hashinaka, K. et al. (1988) Biochemistry 27:5906.
- 10. Hansson, M. et al. (2006) Arch. Biochem. Biophys. 445:214.
- 11. Nauseef, W.M. et al. (1992) Blood 80:2622.
- 12. Tiruppathi, C. et al. (2004) Proc. Natl. Acad. Sci. 101:7699.