

## Mouse MCP-1 Fast ELISA Kit

Catalog Number: EA102912

### Assay Principle

The OriGene Mouse Ccl2 Pre-Coated ELISA (Enzyme-Linked Immunosorbent Assay) kit is a solid phase immunoassay specially designed to measure Mouse Ccl2 with a 96-well strip plate that is pre-coated with antibody specific for Ccl2. The detection antibody is a biotinylated antibody specific for Ccl2. The capture antibody is monoclonal antibody from rat, the detection antibody is polyclonal antibody from goat. The kit contains recombinant Mouse Ccl2 with immunogen: Expression system for standard: E.coli; Immunogen sequence: Q24-R96. The kit is analytically validated with ready to use reagents.

To measure Mouse Ccl2, add standards and samples to the wells, then add the biotinylated detection antibody. Wash the wells with PBS or TBS buffer, and add Avidin-Biotin-Peroxidase Complex (ABC-HRP). Wash away the unbounded ABC-HRP with PBS or TBS buffer and add TMB. TMB is substrate to HRP and will be catalyzed to produce a blue color product, which changes into yellow after adding acidic stop solution. The density of the yellow product is linearly proportional to Mouse Ccl2 in the sample. Read the density of the yellow product in each well using a plate reader, and benchmark the sample wells' readings against the standard curve to determine the concentration of Mouse Ccl2 in the sample.

### Overview

<b>Product Name</b>	Mouse MCP-1 Fast ELISA Kit
<b>Reactive Species</b>	Mouse
<b>Size</b>	96wells/kit, with removable strips.
<b>Description</b>	The Fast version of ELISA kits, assay takes less than 1.5 hours. Detect Mouse Mcp1/CCL2 with < 6pg/ml sensitivity. Format: 96-well plate with removable strips. Compatible samples: cell culture supernates and serum. This is a TMB colorimetric sandwich ELISA kit with short assay time and fast experiment set up. Mcp1/CCL2 tissue specificity:
<b>Sensitivity</b>	< 6pg/ml *The sensitivity or the minimum detectable dose (MDD) is the lower limit of target protein that can be detected by the kit. It is determined by adding two standard deviations to the mean O.D. value of twenty (20) blank wells and calculating the corresponding concentration.
<b>Detection Range</b>	15.6pg/ml-1000pg/ml
<b>Storage Instructions</b>	Store at 4°C for 6 months, at -20°C for 12 months. Avoid multiple freeze-thaw cycles (Shipped with wet ice.)
<b>Uniprot ID</b>	P10148

## Technical Details

<b>Capture/Detection Antibodies</b>	<i>The capture antibody is monoclonal antibody from rat, the detection antibody is polyclonal antibody from goat.</i>
<b>Specificity</b>	<i>Natural and recombinant Mouse Ccl2</i>
<b>Immunogen</b>	<i>Expression system for standard: E.coli; Immunogen sequence: Q24-R96</i>
<b>Cross Reactivity</b>	<i>There is no detectable cross-reactivity with other relevant proteins.</i>

## Notice Before Application

Please read the following instructions before starting the experiment.

1. To inspect the validity of experiment operation and the appropriateness of sample dilution proportion, pilot experiment using standards and a small number of samples is recommended.
2. Before using the Kit, spin tubes and bring down all components to the bottom of tubes.
3. Don't let 96-well plate dry, for dry plate will inactivate active components on plate.
4. Don't reuse tips and tubes to avoid cross contamination.
5. Avoid using the reagents from different batches together.

## Kit Components/Materials Provided

<b>Description</b>	<b>Quantity</b>	<b>Volume</b>
<b>Anti-Mouse Ccl2 Pre-coated 96-well strip microplate</b>	<b>1</b>	<b>12 strips of 8 wells</b>
<b>Mouse Ccl2 Standard</b>	<b>2</b>	<b>10ng/tube</b>
<b>Mouse Ccl2 Biotinylated antibody (50x)</b>	<b>1</b>	<b>130 µl</b>
<b>Avidin-Biotin-Peroxidase Complex (30x)</b>	<b>1</b>	<b>400 µl</b>
<b>Sample Diluent</b>	<b>1</b>	<b>30ml</b>
<b>Antibody Diluent</b>	<b>1</b>	<b>12ml</b>
<b>Avidin-Biotin-Peroxidase Diluent</b>	<b>1</b>	<b>12ml</b>
<b>Color Developing Reagent (TMB)</b>	<b>1</b>	<b>10ml</b>
<b>Stop Solution</b>	<b>1</b>	<b>10ml</b>

Plate Sealers

4

Piece

## Required Materials That Are Not Supplied

Microplate Reader capable of reading absorbance at 450nm.

Automated plate washer (optional)

Pipettes and pipette tips capable of precisely dispensing 0.5 µl through 1 ml volumes of aqueous solutions.

Multichannel pipettes are recommended for large amount of samples.

Deionized or distilled water.

500ml graduated cylinders.

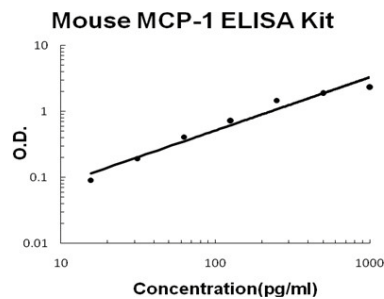
Test tubes for dilution.

## Mouse MCP-1 Fast ELISA Kit (EA102912) Standard Curve Example

Highest O.D. value might be higher or lower than in the example. The experiment result is statistically significant if the highest O.D. value is no less than 1.0.

Concentration 0 (pg/ml)	15.6	31.2	62.5	125	250	500	1000
O.D.	0.014	0.090	0.193	0.406	0.726	1.465	2.338

Mouse MCP-1 ELISA Kit standard curve



A standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.

## Intra/Inter Assay Variability

OriGene spend great efforts in documenting lot to lot variability and make sure our assay kits produce robust data that are reproducible.

**Intra-Assay Precision (Precision within an assay):** Three samples of known concentration were tested on one plate to assess intra-assay precision.

**Inter-Assay Precision (Precision across assays):** Three samples of known concentration were tested in separate assays to assess inter-assay precision.

Sample	Intra-Assay Precision			Inter-Assay Precision		
	1	2	3	1	2	3
n	16	16	16	24	24	24
Mean (pg/ml)	39	224	394	40	231	398
Standard deviation	1.56	10.75	27.18	2	12.47	27.46
CV (%)	4%	4.8%	6.9%	5%	5.4%	6.9%

## Reproducibility

To assay reproducibility, three samples with differing target protein concentrations were assayed using four different lots.

Lots	Lot1 (pg/ml)	Lot2 (pg/ml)	Lot3 (pg/ml)	Lot4 (pg/ml)	Mean (pg/ml)	Standard Deviation	CV (%)
Sample 1	39	36	34	34	35	2.04	5.8%
Sample 2	224	228	228	219	224	3.69	1.6%
Sample 3	394	375	431	394	398	20.3	5.1%

\*number of samples for each test n=16.

## Preparation Before The Experiment

Item	Preparation
All reagents	Bring all reagents to 37°C prior to use. Also the TMB incubation time estimate (20-25min) is based on 37°C.
Wash buffer	Dissolve the included powder in 1000ml of deionized water. Excess wash buffer can be stored for up to one week at 4°C.
Biotinylated Anti-Mouse Ccl2 antibody	It is recommended to prepare this reagent immediately prior to use by diluting the Human ANGPT1 Biotinylated antibody (50x) 1:50 with Antibody Diluent. Prepare 50 µl by adding 1 µl of Biotinylated antibody (50x) to 49 µl of Antibody Diluent. Mix gently and thoroughly and use within 2 hours of generation.
Avidin-Biotin-Peroxidase Complex	It is recommended to prepare this reagent immediately prior to use by diluting the Avidin-Biotin-Peroxidase Complex (30x) 1:30 with Avidin-Biotin-Peroxidase Diluent. Prepare 300 µl by adding 10 µl of Avidin-Biotin-Peroxidase Complex (30x) to 290 µl of Avidin-Biotin-Peroxidase Diluent. Mix gently and thoroughly and use within 2 hours of generation.

Mouse Ccl2 Standard	It is recommended that the standards be prepared no more than 2 hours prior to performing the experiment. Use one 10ng of lyophilized Mouse Ccl2 standard for each experiment. Gently spin the vial prior to use. Reconstitute the standard to a stock concentration of 10ng/ml using 1ml of sample diluent. Allow the standard to sit for a minimum of 10 minutes with gentle agitation prior to making dilutions.
Microplate	The included microplate is coated with capture antibodies and ready-to-use. It does not require additional washing or blocking. The unused well strips should be sealed and stored in the original packaging.

## Dilution of Mouse Ccl2 Standard

1. Number tubes 1-8. Final Concentrations to be Tube # 1 – 1000pg/ml, #2 – 500pg/ml, #3 – 250pg/ml, #4 – 125pg/ml, #5 – 62.5pg/ml, #6 – 31.25pg/ml, #7 – 15.625pg/ml, #8 – 0.0 (Blank).
2. To generate standard #1, add 100µl of the reconstituted standard stock solution of 10ng/ml and 900µl of sample diluent to tube #1 for a final volume of 1000µl. Mix thoroughly.
3. Add 300 µl of sample diluent to tubes # 2-7.
4. To generate standard #2, add 300 µl of standard #1 from tube #1 to tube #2 for a final volume of 600 µl. Mix thoroughly.
5. To generate standard #3, add 300 µl of standard #2 from tube #2 to tube #3 for a final volume of 600 µl. Mix thoroughly.
6. Continue the serial dilution for tube #4-7.
7. Tube #8 is a blank standard to be used with every experiment.

## Sample Preparation and Storage

These sample collection instructions and storage conditions are intended as a general guideline and the sample stability has not been evaluated.

Sample Type	Procedure
Cell culture supernatants	Clear sample of particulates by centrifugation, assay immediately or store samples at -20°C.
Serum	Use a serum separator tube (SST) and allow serum to clot at room temperature for about four hours. Then, centrifuge for 15 min at approximately 1,000 xg. assay immediately or store samples at -20°C.

## Sample Dilution

The target protein concentration should be estimated and appropriate sample dilutions should be selected such that the final protein concentration lies near the middle of the linear dynamic range of the assay.

It is recommended to prepare 150 µl of sample for each replicate to be assayed. The samples should be diluted with sample diluent and mixed gently.

## Assay protocol

*It is recommended that all reagents and materials be equilibrated to 37°C/room temperature prior to the experiment (see Preparation Before The Experiment if you have missed this information).*

1. Prepare all reagents and working standards as directed previously.
2. Remove excess microplate strips from the plate frame and seal and store them in the original packaging.
3. Add 50 µl of the standard, samples, or control per well. And add 50 µl of the prepared 1x Biotinylated Anti-Human ANGPT1 antibody per well. Add 50 µl of the sample diluent buffer and 50 µl of the prepared 1x Biotinylated Anti-Human ANGPT1 antibody into the control well (Zero well). At least two replicates of each standard, sample, or control is recommended.
4. Cover with the plate sealer provided and incubate for 60 minutes at RT.
5. Wash the plate 3 times with the 1x wash buffer.
  - a. Discard the liquid in the wells into an appropriate waste receptacle. Then, invert the plate on the benchtop onto a paper towel and tap the plate to gently blot any remaining liquid. It is recommended that the wells are not allowed to completely dry at any time.
  - b. Add 300 µl of the 1x wash buffer to each assay well. (For cleaner background incubate for 60 seconds between each wash).
  - c. Repeat steps a-b 2 additional times.
6. Add 100 µl of the prepared 1x Avidin-Biotin-Peroxidase Complex into each well. Cover with plate sealer provided and incubate for 15 minutes at RT.
7. Wash the plate 5 times with the 1x wash buffer.
  - a. Discard the liquid in the wells into an appropriate waste receptacle. Then, invert the plate on the benchtop onto a paper towel and tap the plate to gently blot any remaining liquid. It is recommended that the wells are not allowed to completely dry at any time.
  - b. Add 300 µl of the 1x wash buffer to each assay well. (For cleaner background incubate for 60 seconds between each wash).
  - c. Repeat steps a-b 4 additional times.
8. Add 90 µl of Color Developing Reagent to each well and incubate in the dark for 30 minutes at RT (or 25-30 minutes at 37°C). (The optimal incubation time must be empirically determined. A guideline to look for is blue shading the top four standard wells, while the remaining standards remain clear.)
9. Add 100 µl of Stop Solution to each well. The color should immediately change to yellow.
10. Within 30 minutes of stopping the reaction, the O.D. absorbance should be read with a microplate reader at 450nm.

## **Data Analysis**

*Average the duplicate readings for each standard, sample, and control. Subtract the average zero standard O.D. reading.*

*It is recommended that a standard curve be created using computer software to generate a four parameter logistic (4-PL) curve-fit. A free program capable of generating a four parameter logistic (4-PL) curve-fit can be found online at: [www.myassays.com/four-parameter-logistic-curve.assay](http://www.myassays.com/four-parameter-logistic-curve.assay).*

*Alternatively, plot the mean absorbance for each standard against the concentration. The measured concentration in the sample can be interpolated by using linear regression of each average relative OD against the standard curve generated using curve fitting software. This will generate an adequate but less precise fit of the data.*

*For diluted samples, the concentration reading from the standard curve must be multiplied by the dilution factor.*

## **Background on Ccl2**



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*Monocyte chemotactic protein-1 (MCP-1) is a member of the small inducible gene (SIG) family. It has been shown to play a role in the recruitment of monocytes to sites of injury and infection. By analysis of a panel of somatic cell hybrids, we have localized the MCP-1 gene, designated SCYA2, to human chromosome 17. In situ hybridization confirmed this assignment and further localized the gene to 17q11.2-q21.1. MCP-1 plays a unique and crucial role in the initiation of atherosclerosis and may provide a new therapeutic target in this disorder. Monocyte chemoattractant protein-1 (MCP-1), regulated on activation, normal T cell expressed and secreted, and stimulation of monocytes from healthy carriers of the genotype GG with Mycobacterium tuberculosis antigens yielded higher MCP-1. The standard used in this kit is recombinant mouse MCP-1 (Q24-R96) with the molecular mass of 8.5Kda.*