

## Horse equine TGF-Beta 3 ELISA Kit

Catalog Number: EA102862

### Assay Principle

The OriGene Equine TGFB3 Pre-Coated ELISA (Enzyme-Linked Immunosorbent Assay) kit is a solid phase immunoassay specially designed to measure Equine TGFB3 with a 96-well strip plate that is pre-coated with antibody specific for TGFB3. The detection antibody is a biotinylated antibody specific for TGFB3. The capture antibody is monoclonal antibody from mouse, the detection antibody is polyclonal antibody from goat. The kit contains recombinant Equine TGFB3 with immunogen: Expression system for standard: sf21; Immunogen sequence: A301-S412. The kit is analytically validated with ready to use reagents.

To measure Equine TGFB3, add standards and samples to the wells, then add the biotinylated detection antibody. Wash the wells with PBS or TBS buffer, and add Avidin-Biotin-Peroxidase Complex (ABC-HRP). Wash away the unbound ABC-HRP with PBS or TBS buffer and add TMB. TMB is substrate to HRP and will be catalyzed to produce a blue color product, which changes into yellow after adding acidic stop solution. The density of the yellow product is linearly proportional to Equine TGFB3 in the sample. Read the density of the yellow product in each well using a plate reader, and benchmark the sample wells' readings against the standard curve to determine the concentration of Equine TGFB3 in the sample.

### Overview

<b>Product Name</b>	Horse equine TGF-Beta 3 ELISA Kit
<b>Reactive Species</b>	Equine
<b>Size</b>	96wells/kit, with removable strips.
<b>Description</b>	Sandwich High Sensitivity ELISA kit for Quantitative Detection of activated horse equine TGF-beta 3. 96wells/kit, with removable strips.
<b>Sensitivity</b>	< 10pg/ml *The sensitivity or the minimum detectable dose (MDD) is the lower limit of target protein that can be detected by the kit. It is determined by adding two standard deviations to the mean O.D. value of twenty (20) blank wells and calculating the corresponding concentration.
<b>Detection Range</b>	31.2pg/ml-2000pg/ml
<b>Storage Instructions</b>	Store at 4 °C for 6 months, at -20 °C for 12 months. Avoid multiple freeze-thaw cycles (Shipped with wet ice.)
<b>Uniprot ID</b>	P10600

## Technical Details

<b>Capture/Detection Antibodies</b>	The capture antibody is monoclonal antibody from mouse, the detection antibody is polyclonal antibody from goat.
<b>Specificity</b>	Natural and recombinant Equine TGFB3
<b>Immunogen</b>	Expression system for standard: sf21; Immunogen sequence: A301-S412
<b>Cross Reactivity</b>	There is no detectable cross-reactivity with other relevant proteins.

## Notice Before Application

Please read the following instructions before starting the experiment.

1. To inspect the validity of experiment operation and the appropriateness of sample dilution proportion, pilot experiment using standards and a small number of samples is recommended.
2. Before using the Kit, spin tubes and bring down all components to the bottom of tubes.
3. Don't let 96-well plate dry, for dry plate will inactivate active components on plate.
4. Don't reuse tips and tubes to avoid cross contamination.
5. Avoid using the reagents from different batches together.

## Kit Components/Materials Provided

Description	Quantity	Volume
Anti-Equine TGFB3 Pre-coated 96-well strip microplate	1	12 strips of 8 wells
Equine TGFB3 Standard	2	10ng/tube
Equine TGFB3 Biotinylated antibody (100x)	1	130 µl
Avidin-Biotin-Peroxidase Complex (100x)	1	130 µl
Sample Diluent	1	30ml
Antibody Diluent	1	12ml
Avidin-Biotin-Peroxidase Diluent	1	12ml
Color Developing Reagent (TMB)	1	10ml
Stop Solution	1	10ml
Plate Sealers	4	Piece

\*Why there is no wash buffer? Our Avidin-Biotin-Peroxidase Diluent contains the detergent (TWEEN) normally present in other companies' ELISA kits. This saves you the step of having to wash with the special wash buffer and achieve similar or better signal to noise ratio. The wash can use regular wash buffers (PBS, TBS etc.) commonly found in labs.

## Required Materials That Are Not Supplied

Microplate Reader capable of reading absorbance at 450nm.

Automated plate washer (optional)

1000ml of 1X wash buffer (TBS or PBS)

Pipettes and pipette tips capable of precisely dispensing 0.5 µl through 1 ml volumes of aqueous solutions.

Multichannel pipettes are recommended for large amount of samples.

Deionized or distilled water.

500ml graduated cylinders.

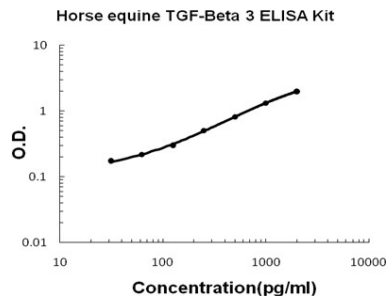
Test tubes for dilution.

## Horse equine TGF-Beta 3 ELISA Kit (EA102862) Standard Curve Example

Highest O.D. value might be higher or lower than in the example. The experiment result is statistically significant if the highest O.D. value is no less than 1.0.

<b>Concentration 0</b> (pg/ml)	<b>2000</b>	<b>2000</b>	<b>2000</b>	<b>2000</b>	<b>2000</b>	<b>2000</b>	<b>2000</b>	<b>2000</b>
<b>O.D.</b>	<b>0.092</b>	<b>0.175</b>	<b>0.217</b>	<b>0.302</b>	<b>0.504</b>	<b>0.812</b>	<b>1.321</b>	<b>1.984</b>

**Horse equine TGF-beta 3 ELISA Kit standard curve**



A standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.

## Intra/Inter Assay Variability

OriGene spend great efforts in documenting lot to lot variability and make sure our assay kits produce robust data that are reproducible.

**Intra-Assay Precision (Precision within an assay):** Three samples of known concentration were tested on one plate to assess intra-assay precision.

**Inter-Assay Precision (Precision across assays):** Three samples of known concentration were tested in separate assays to assess inter-assay

precision.

Sample	Intra-Assay Precision			Inter-Assay Precision		
	1	2	3	1	2	3
n	16	16	16	24	24	24
Mean(pg/ml)	271	631	1153	294	670	1265
Standard deviation	14.4	29	45	18.82	36.4	72.85
CV(%)	5.3	4.6	3.9	6.4	5.7	5.8

## Preparation Before The Experiment

Item	Preparation
All reagents	Bring all reagents to 37°C prior to use. The assay can also be done at room temperature however we recommend doing it at 37°C for best consistency with our QC results. Also the TMB incubation time estimate (15-25min) is based on 37°C.
Wash buffer	Prepare 1000ml of 1X PBS or TBS for wash buffer.
Biotinylated Anti-Equine TGFB3 antibody	It is recommended to prepare this reagent immediately prior to use by diluting the Equine TGFB3 Biotinylated antibody (100x) 1: 100 with Antibody Diluent. Prepare 100 µl by adding 1 µl of Biotinylated antibody (100x) to 99 µl of Antibody Diluent for each well. Mix gently and thoroughly and use within 2 hours of generation.
Avidin-Biotin-Peroxidase Complex	It is recommended to prepare this reagent immediately prior to use by diluting the Avidin-Biotin-Peroxidase Complex (100x) 1: 100 with Avidin-Biotin-Peroxidase Diluent. Prepare 100 µl by adding 1 µl of Avidin-Biotin-Peroxidase Complex (100x) to 99 µl of Avidin-Biotin-Peroxidase Diluent for each well. Mix gently and thoroughly and use within 2 hours of generation.
Equine TGFB3 Standard	It is recommended that the standards be prepared no more than 2 hours prior to performing the experiment. Use one 10ng of lyophilized Equine TGFB3 standard for each experiment. Gently spin the vial prior to use. Reconstitute the standard to a stock concentration of 10ng/ml using 1ml of sample diluent. Allow the standard to sit for a minimum of 10 minutes with gentle agitation prior to making dilutions.
Microplate	The included microplate is coated with capture antibodies and ready-to-use. It does not require additional washing or blocking. The unused well strips should be sealed and stored in the original packaging.

## Dilution of Equine TGFB3 Standard

1. Number tubes 1-8. Final Concentrations to be Tube # 1 – 2000pg/ml, #2 – 1000pg/ml, #3 – 500pg/ml, #4 – 250pg/ml, #5 – 125pg/ml, #6 –

62.5pg/ml, #7 – 31.25pg/ml, #8 – 0.0 (Blank).

2. To generate standard #1, add 200µl of the reconstituted standard stock solution of 10ng/ml and 800µl of sample diluent to tube #1 for a final volume of 1000µl. Mix thoroughly.
3. Add 300 µl of sample diluent to tubes # 2-7.
4. To generate standard #2, add 300 µl of standard #1 from tube #1 to tube #2 for a final volume of 600 µl. Mix thoroughly.
5. To generate standard #3, add 300 µl of standard #2 from tube #2 to tube #3 for a final volume of 600 µl. Mix thoroughly.
6. Continue the serial dilution for tube #4-7.
7. Tube #8 is a blank standard to be used with every experiment.

## Sample Preparation and Storage

These sample collection instructions and storage conditions are intended as a general guideline and the sample stability has not been evaluated.

Sample Type	Procedure
Cell culture supernatants	Clear sample of particulates by centrifugation, assay immediately or store samples at -20°C.
Serum	Use a serum separator tube (SST) and allow serum to clot at room temperature for about four hours. Then, centrifuge for 15 min at approximately 1,000 x g. assay immediately or store samples at -20°C.
Plasma	Collect plasma using EDTA as an anticoagulant. Centrifuge for 15 min at approximately 1,000 x g. Assay immediately or store samples at -20°C. <i>*Note: it is important to not use anticoagulants other than the ones described above to treat plasma for other anticoagulants could block the antibody binding site.</i>
Urine	Collect the first urine of the day, micturate directly into a sterile container. Remove impurities by centrifugation, assay immediately or aliquot and store samples at -20°C.

*\*Note: To detect TGFB3 in samples, you need to activate TGFB3 in samples prior to the assay.*

TGF beta 3 is mostly contained as inactive form in samples, please activate it before assay. Don't activate recombinant TGF beta 3.

**Solution A:** 1N HCl: add 8.33ml of 12N HCl into 91.67ml of H<sub>2</sub>O.

**Solution B:** 1.2N NaOH/0.5M HEPES: add 12ml of 10N NaOH and 11.9g HEPES into 75ml of H<sub>2</sub>O, add H<sub>2</sub>O to adjust volume to 100ml.

### Activate the sample

Cell culture supernates, urine: add activating reagent pro rata, i.e. add 20µl of Solution A into 100µl of sample, 10 min later, add 20µl of Solution B. PH 7.0-7.6.

Serum, plasma (EDTA): add activating reagent pro rata, i.e. add 20µl of Solution A into 40µl of sample, 10 min later, add 20µl of Solution B. PH 7.0-7.6.

*It is unnecessary to activate the recombinant TGF-beta 3.*

*Sample was diluted partly after adding activating reagent, so please pay attention to this when calculate target protein concentration.*

## **Sample Dilution**

*The target protein concentration should be estimated and appropriate sample dilutions should be selected such that the final protein concentration lies near the middle of the linear dynamic range of the assay.*

*It is recommended to prepare 150 µl of sample for each replicate to be assayed. The samples should be diluted with sample diluent and mixed gently.*

## **Assay protocol**

*It is recommended that all reagents and materials be equilibrated to 37°C/room temperature prior to the experiment (see Preparation Before The Experiment if you have missed this information).*

- 1. Prepare all reagents and working standards as directed previously.*
- 2. Remove excess microplate strips from the plate frame and seal and store them in the original packaging.*
- 3. Add 100 µl of the standard, samples, or control per well. Add 100 µl of the sample diluent buffer into the control well (Zero well). At least two replicates of each standard, sample, or control is recommended.*
- 4. Cover with the plate sealer provided and incubate for 120 minutes at RT (or 90 min. at 37 °C).*
- 5. Remove the cover and discard the liquid in the wells into an appropriate waste receptacle. Invert the plate on the benchtop onto a paper towel and tap the plate to gently blot any remaining liquid. It is recommended that the wells are not allowed to completely dry at any time.*
- 6. Add 100 µl of the prepared 1x Biotinylated Anti-Equine TGFB3 antibody to each well.*
- 7. Cover with plate sealer and incubate for 90 minutes at RT (or 60 minutes at 37°C).*
- 8. Wash the plate 3 times with the 1x wash buffer.*
  - a. Discard the liquid in the wells into an appropriate waste receptacle. Then, invert the plate on the benchtop onto a paper towel and tap the plate to gently blot any remaining liquid. It is recommended that the wells are not allowed to completely dry at any time.*
  - b. Add 300 µl of the 1x wash buffer to each assay well. (For cleaner background incubate for 60 seconds between each wash).*
  - c. Repeat steps a-b 2 additional times.*
- 9. Add 100 µl of the prepared 1x Avidin-Biotin-Peroxidase Complex into each well. Cover with the plate sealer provided and incubate for 40 minutes at RT (or 30 minutes at 37°C).*
- 10. Wash the plate 5 times with the 1x wash buffer.*
  - a. Discard the liquid in the wells into an appropriate waste receptacle. Then, invert the plate on the benchtop onto a paper towel and tap the plate to gently blot any remaining liquid. It is recommended that the wells are not allowed to completely dry at any time.*
  - b. Add 300 µl of the 1x wash buffer to each assay well. (For cleaner background incubate for 60 seconds between each wash).*
  - c. Repeat steps a-b 4 additional times.*
- 11. Add 90 µl of Color Developing Reagent to each well. Cover with the plate sealer provided and incubate in the dark for 30 minutes at RT (or 15-25 minutes at 37°C). (The optimal incubation time must be empirically determined. A guideline to look for is blue shading the top four standard wells, while the remaining standards remain clear.)*
- 12. Add 100 µl of Stop Solution to each well. The color should immediately change to yellow.*

13. Within 30 minutes of stopping the reaction, the O.D. absorbance should be read with a microplate reader at 450nm.

## Data Analysis

Average the duplicate readings for each standard, sample, and control. Subtract the average zero standard O.D. reading.

It is recommended that a standard curve be created using computer software to generate a four parameter logistic (4-PL) curve-fit. A free program capable of generating a four parameter logistic (4-PL) curve-fit can be found online at: [www.myassays.com/four-parameter-logistic-curve.assay](http://www.myassays.com/four-parameter-logistic-curve.assay).

Alternatively, plot the mean absorbance for each standard against the concentration. The measured concentration in the sample can be interpolated by using linear regression of each average relative OD against the standard curve generated using curve fitting software. This will generate an adequate but less precise fit of the data.

For diluted samples, the concentration reading from the standard curve must be multiplied by the dilution factor.

## Background on TGFB3

Transforming growth factor beta 3 (TGF-beta 3) is a type of protein, known as a cytokine, which is involved in cell differentiation, embryogenesis and development. It belongs to a large family of cytokines called the Transforming growth factor beta superfamily. TGF-beta 3 is believed to regulate molecules involved in cellular adhesion and extracellular matrix (ECM) formation during the process of palate development. Without TGF-beta 3, mammals develop a deformity known as a cleft palate. This is caused by failure of epithelial cells in both sides of the developing palate to fuse. TGF-beta 3 also plays an essential role in controlling the development of lungs in mammals, by also regulating cell adhesion and ECM formation in this tissue, and controls wound healing by regulating the movements of epidermal and dermal cells in injured skin. TGF-beta 3 activated Lef1 in the absence of beta-catenin (CTNNB1) via nuclear phospho-Smad2 and Smad4.