

## Human CD5L / CT 2 ELISA Kit

**Catalog Number:**EA102293

### Assay Principle

The OriGene Human CD5L Pre-Coated ELISA (Enzyme-Linked Immunosorbent Assay) kit is a solid phase immunoassay specially designed to measure Human CD5L with a 96-well strip plate that is pre-coated with antibody specific for CD5L. The detection antibody is a biotinylated antibody specific for CD5L. The capture antibody is monoclonal antibody from mouse, the detection antibody is polyclonal antibody from goat. The kit contains recombinant Human CD5L with immunogen: Expression system for standard: NSO; Immunogen sequence: S20-G347. The kit is analytically validated with ready to use reagents.

To measure Human CD5L, add standards and samples to the wells, then add the biotinylated detection antibody. Wash the wells with PBS or TBS buffer, and add Avidin-Biotin-Peroxidase Complex (ABC-HRP). Wash away the unbounded ABC-HRP with PBS or TBS buffer and add TMB. TMB is substrate to HRP and will be catalyzed to produce a blue color product, which changes into yellow after adding acidic stop solution. The density of the yellow product is linearly proportional to Human CD5L in the sample. Read the density of the yellow product in each well using a plate reader, and benchmark the sample wells' readings against the standard curve to determine the concentration of Human CD5L in the sample.

### Overview

<b>Product Name</b>	Human CD5L / CT 2 ELISA Kit
<b>Reactive Species</b>	Human
<b>Size</b>	96wells/kit, with removable strips.
<b>Description</b>	Sandwich High Sensitivity ELISA kit for Quantitative Detection of activated Human CD5L. 96wells/kit, with removable strips.
<b>Sensitivity</b>	<10pg/ml *The sensitivity or the minimum detectable dose (MDD) is the lower limit of target protein that can be detected by the kit. It is determined by adding two standard deviations to the mean O.D. value of twenty (20) blank wells and calculating the corresponding concentration.
<b>Detection Range</b>	156pg/ml-10000pg/ml
<b>Storage Instructions</b>	Store at 4°C for 6 months, at -20°C for 12 months. Avoid multiple freeze-thaw cycles(Shipped with wet ice.)
<b>Uniprot ID</b>	O43866

## Technical Details

<b>Capture/Detection Antibodies</b>	The capture antibody is monoclonal antibody from mouse, the detection antibody is polyclonal antibody from goat.
<b>Specificity</b>	Natural and recombinant Human CD5L
<b>Immunogen</b>	Expression system for standard: NSO; Immunogen sequence: S20-G347
<b>Cross Reactivity</b>	There is no detectable cross-reactivity with other relevant proteins.

## Notice Before Application

Please read the following instructions before starting the experiment.

1. To inspect the validity of experiment operation and the appropriateness of sample dilution proportion, pilot experiment using standards and a small number of samples is recommended.
2. Before using the Kit, spin tubes and bring down all components to the bottom of tubes.
3. Don't let 96-well plate dry, for dry plate will inactivate active components on plate.
4. Don't reuse tips and tubes to avoid cross contamination.
5. Avoid using the reagents from different batches together.

## Kit Components/Materials Provided

Description	Quantity	Volume
Anti-Human CD5L Pre-coated 96-well strip microplate	1	12 strips of 8 wells
Human CD5L Standard	2	10ng/tube
Human CD5L Biotinylated antibody (100x)	1	130 µl
Avidin-Biotin-Peroxidase Complex (100x)	1	130 µl
Sample Diluent	1	30ml
Antibody Diluent	1	12ml
Avidin-Biotin-Peroxidase Diluent	1	12ml
Color Developing Reagent (TMB)	1	10ml
Stop Solution	1	10ml
Plate Sealers	4	Piece

\*Why there is no wash buffer? Our Avidin-Biotin-Peroxidase Diluent contains the detergent (TWEEN) normally present in other companies' ELISA kits. This saves you the step of having to wash with the special wash buffer and achieve similar or better signal to noise ratio. The wash can use regular wash buffers (PBS, TBS etc.) commonly found in labs.

## Required Materials That Are Not Supplied

Microplate Reader capable of reading absorbance at 450nm.

Automated plate washer (optional)

1000ml of 1X wash buffer (TBS or PBS)

Pipettes and pipette tips capable of precisely dispensing 0.5 µl through 1 ml volumes of aqueous solutions.

Multichannel pipettes are recommended for large amount of samples.

Deionized or distilled water.

500ml graduated cylinders.

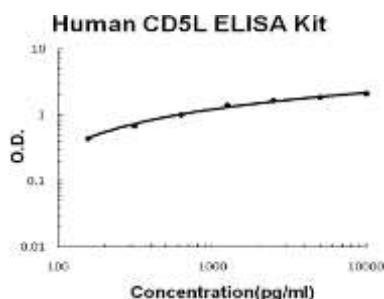
Test tubes for dilution.

## Human CD5L / CT 2 ELISA Kit (EA102293) Standard Curve Example

Highest O.D. value might be higher or lower than in the example. The experiment result is statistically significant if the highest O.D. value is no less than 1.0.

Concentration 0 (pg/ml)	156	312	625	1250	2500	5000	10000	
O.D.	0.008	0.433	0.677	1.005	1.409	1.650	1.849	2.099

### Human CD5L ELISA Kit standard curve



A standard curve is provided for demonstration only. A standard curve should be generated for each set of samples assayed.

## Intra/Inter Assay Variability

OriGene spend great efforts in documenting lot to lot variability and make sure our assay kits produce robust data that are reproducible.

**Intra-Assay Precision (Precision within an assay):** Three samples of known concentration were tested on one plate to assess intra-assay precision.

**Inter-Assay Precision (Precision across assays):** Three samples of known concentration were tested in separate assays to assess inter-assay

precision.

#### Intra-Assay Precision

Sample	1	2	3
n	16	16	16
Mean(pg/ml)	245	1172	5450
Standard deviation	10.04	79.69	414.2
CV(%)	4.1%	6.8%	7.6%

#### Inter-Assay Precision

	1	2	3
n	24	24	24
Mean	224	1288	5192
Standard deviation	13.21	91.44	415.36
CV(%)	5.9%	7.1%	8%

## Reproducibility

To assay reproducibility, three samples with differing target protein concentrations were assayed using four different lots.

Lots	Lot1 (pg/ml)	Lot2 (pg/ml)	Lot3 (pg/ml)	Lot4 (pg/ml)	Mean (pg/ml)	Standard Deviation	CV (%)
Sample 1	245	242	250	244	245	2.94	1.2%
Sample 2	1172	1254	1148	1193	1191	39.3	3.2%
Sample 3	5450	5193	4994	5419	5264	184.77	3.5%

\*number of samples for each test n=16.

## Preparation Before The Experiment

Item	Preparation
All reagents	Bring all reagents to 37°C prior to use. The assay can also be done at room temperature however we recommend doing it at 37°C for best consistency with our QC results. Also the TMB incubation time estimate (15-25min) is based on 37°C.
Wash buffer	Prepare 1000ml of 1X PBS or TBS for wash buffer.
Biotinylated Anti-Human CD5L antibody	It is recommended to prepare this reagent immediately prior to use by diluting the Human CD5L Biotinylated antibody (100x) 1:100 with Antibody Diluent. Prepare 100 µl by adding 1 µl of Biotinylated antibody (100x) to 99 µl of Antibody Diluent for each well. Mix gently and thoroughly and use within 2 hours of generation.
Avidin-Biotin-Peroxidase Complex	It is recommended to prepare this reagent immediately prior to use by diluting the Avidin-Biotin-Peroxidase Complex (100x) 1:100 with Avidin-Biotin-Peroxidase Diluent. Prepare 100 µl by adding 1 µl of Avidin-Biotin-Peroxidase Complex (100x) to 99 µl of Avidin-Biotin-Peroxidase Diluent for each well. Mix gently and thoroughly and use within 2 hours of generation.
Human CD5L Standard	It is recommended that the standards be prepared no more than 2 hours prior to performing the

	experiment. Use one 10ng of lyophilized Human CD5L standard for each experiment. Gently spin the vial prior to use. Reconstitute the standard to a stock concentration of 10ng/ml using 1ml of sample diluent. Allow the standard to sit for a minimum of 10 minutes with gentle agitation prior to making dilutions.
Microplate	The included microplate is coated with capture antibodies and ready-to-use. It does not require additional washing or blocking. The unused well strips should be sealed and stored in the original packaging.

## Dilution of Human CD5L Standard

1. Number tubes 1-8. Final Concentrations to be Tube # 1 – 10000pg/ml, #2 – 5000pg/ml, #3 – 2500pg/ml, #4 – 1250pg/ml, #5 – 625pg/ml, #6 – 312.5pg/ml, #7 – 156.25pg/ml, #8 – 0.0 (Blank).
2. For standard #1, add 1000µl of undiluted standard stock solution to tube #1.
3. Add 300 µl of sample diluent to tubes # 2-7.
4. To generate standard #2, add 300 µl of standard #1 from tube #1 to tube #2 for a final volume of 600 µl. Mix thoroughly.
5. To generate standard #3, add 300 µl of standard #2 from tube #2 to tube #3 for a final volume of 600 µl. Mix thoroughly.
6. Continue the serial dilution for tube #4-7.
7. Tube #8 is a blank standard to be used with every experiment.

## Sample Preparation and Storage

These sample collection instructions and storage conditions are intended as a general guideline and the sample stability has not been evaluated.

Sample Type	Procedure
Cell culture supernatants	Clear sample of particulates by centrifugation, assay immediately or store samples at -20°C.
Serum	Use a serum separator tube (SST) and allow serum to clot at room temperature for about four hours. Then, centrifuge for 15 min at approximately 1,000 x g. assay immediately or store samples at -20°C.
Plasma	Collect plasma using heparin or EDTA as an anticoagulant. Centrifuge for 15 min at approximately 1,000 x g. Assay immediately or store samples at -20°C. *Note: it is important to not use anticoagulants other than the ones described above to treat plasma for other anticoagulants could block the antibody binding site.
Cell lysates	Lyse the cells, make sure there are no visible cell sediments. Centrifuge cell lysates at approximately 10000 X g for 5 min. Collect the supernatant.

**\*Note:** To detect CD5L in samples, you need to activate CD5L in samples prior to the assay.

CD5 combines with its ligand (CD5L) to form complex in samples. Free CD5L needs to be released by activation.

**Solution A:** 1N HCl: add 8.33ml of 12N HCl into 91.67ml of H<sub>2</sub>O.

**Solution B:** 1.2N NaOH/0.5M HEPES: add 12ml of 10N NaOH and 11.9g HEPES into 75ml of H<sub>2</sub>O, add H<sub>2</sub>O to adjust volume to 100ml.

### **Activate the sample**

Cell culture supernate: add activating reagent pro rate, i.e. add 20 $\mu$ l of Solution A into 40 $\mu$ l of sample, 10 min later, add 20 $\mu$ l of Solution B. PH7.0-7.6.

Serum: add activating reagent pro rate, i.e. add 20 $\mu$ l of Solution A into 40 $\mu$ l of sample, 10 min later, add 20 $\mu$ l of Solution B. PH7.0-7.6. Prior to the assay, the activated sample requires a 2000-fold dilution. Add 10 $\mu$ l of activated sample into 390 $\mu$ l wash buffer (1 : 40). And then add 4 $\mu$ l of above diluted reagent into 196 $\mu$ l sample diluent buffer (1 : 50).

Plasma (heparin, EDTA): add activating reagent pro rate, i.e. add 20 $\mu$ l of Solution A into 40 $\mu$ l of sample, 10 min later, add 20 $\mu$ l of Solution B. PH7.0-7.6. Prior to the assay, the activated sample requires a 4000-fold dilution. Add 10 $\mu$ l of activated sample into 790 $\mu$ l wash buffer (1 : 80). And then add 4 $\mu$ l of above diluted reagent into 196 $\mu$ l sample diluent buffer (1 : 50). Optimal dilutions should be determined by end users.

It is unnecessary to activate the recombinant CD5L.

Sample was diluted partly after adding activating reagent, so please pay attention to this when calculate target protein concentration.

## **Sample Dilution**

The target protein concentration should be estimated and appropriate sample dilutions should be selected such that the final protein concentration lies near the middle of the linear dynamic range of the assay.

It is recommended to prepare 150  $\mu$ l of sample for each replicate to be assayed. The samples should be diluted with sample diluent and mixed gently.

## **Assay protocol**

It is recommended that all reagents and materials be equilibrated to 37°C/room temperature prior to the experiment (see Preparation Before The Experiment if you have missed this information).

1. Prepare all reagents and working standards as directed previously.
2. Remove excess microplate strips from the plate frame and seal and store them in the original packaging.
3. Add 100  $\mu$ l of the standard, samples, or control per well. Add 100  $\mu$ l of the sample diluent buffer into the control well (Zero well). At least two replicates of each standard, sample, or control is recommended.
4. Cover with the plate sealer provided and incubate for 120 minutes at RT (or 90 min. at 37 °C).
5. Remove the cover and discard the liquid in the wells into an appropriate waste receptacle. Invert the plate on the benchtop onto a paper towel and tap the plate to gently blot any remaining liquid. It is recommended that the wells are not allowed to completely dry at any time.
6. Add 100  $\mu$ l of the prepared 1x Biotinylated Anti-Human CD5L antibody to each well.
7. Cover with plate sealer and incubate for 90 minutes at RT (or 60 minutes at 37°C).

8. Wash the plate 3 times with the 1x wash buffer.
  - a. Discard the liquid in the wells into an appropriate waste receptacle. Then, invert the plate on the benchtop onto a paper towel and tap the plate to gently blot any remaining liquid. It is recommended that the wells are not allowed to completely dry at any time.
  - b. Add 300  $\mu$ l of the 1x wash buffer to each assay well. (For cleaner background incubate for 60 seconds between each wash).
  - c. Repeat steps a-b 2 additional times.
9. Add 100  $\mu$ l of the prepared 1x Avidin-Biotin-Peroxidase Complex into each well. Cover with the plate sealer provided and incubate for 40 minutes at RT (or 30 minutes at 37°C).
10. Wash the plate 5 times with the 1x wash buffer.
  - a. Discard the liquid in the wells into an appropriate waste receptacle. Then, invert the plate on the benchtop onto a paper towel and tap the plate to gently blot any remaining liquid. It is recommended that the wells are not allowed to completely dry at any time.
  - b. Add 300  $\mu$ l of the 1x wash buffer to each assay well. (For cleaner background incubate for 60 seconds between each wash).
  - c. Repeat steps a-b 4 additional times.
11. Add 90  $\mu$ l of Color Developing Reagent to each well. Cover with the plate sealer provided and incubate in the dark for 30 minutes at RT (or 15-25 minutes at 37°C). (The optimal incubation time must be empirically determined. A guideline to look for is blue shading the top four standard wells, while the remaining standards remain clear.)
12. Add 100  $\mu$ l of Stop Solution to each well. The color should immediately change to yellow.
13. Within 30 minutes of stopping the reaction, the O.D. absorbance should be read with a microplate reader at 450nm.

## Data Analysis

Average the duplicate readings for each standard, sample, and control. Subtract the average zero standard O.D. reading.

It is recommended that a standard curve be created using computer software to generate a four parameter logistic (4-PL) curve-fit. A free program capable of generating a four parameter logistic (4-PL) curve-fit can be found online at: [www.myassays.com/four-parameter-logistic-curve.assay](http://www.myassays.com/four-parameter-logistic-curve.assay).

Alternatively, plot the mean absorbance for each standard against the concentration. The measured concentration in the sample can be interpolated by using linear regression of each average relative OD against the standard curve generated using curve fitting software. This will generate an adequate but less precise fit of the data.

For diluted samples, the concentration reading from the standard curve must be multiplied by the dilution factor.

## Background on CD5L

CD5 antigen-like, also known as Sp $\alpha$  and AIM, is a protein that in humans is encoded by the CD5L gene. It is mapped to 1q21-q23 by fluorescence in situ hybridization. It is found that Aim expression is induced in mouse macrophages in response to loading with highly oxidized low density lipoprotein (oxLDL), and that Aim is expressed in foam cells within atherosclerotic lesions. Both the expression of Aim in lesions and its induction by oxLDL require Lxr/Rxr heterodimers. Aim-null macrophages are highly susceptible to oxLDL-induced apoptosis in vitro and undergo accelerated apoptosis in atherosclerotic lesions in vivo. Double knockout of Aim and Ldlr reduce atherosclerotic lesions. Therefore, it is concluded that AIM expression protects macrophages from apoptosis within atherosclerotic lesions, promoting early lesion development.



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