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Human sCD25 (IL-2R) ELISA KIT

1. Intended use

The OriGene sCD25 ELISA kit is a solid phase sandwich ELISA for the *in-vitro* qualitative and quantitative determination of sCD25 in supernatants, buffered solutions or serum and plasma samples. This assay will recognise both natural and recombinant human sCD25.

This kit has been configured research use only. Not suitable for use in therapeutic procedures.

2. Introduction

2.1. Summary

IL-2, one of the most important factors in the human immune system, is a potent T-cell growth factor whose major function is the activation of many cells of the immune system including T-cells, B-cells, macrophages and NK cells. These potent actions are mediated by IL-2 binding and signalling through its associated cell surface receptor IL-2R. This receptor is not expressed on normal or unstimulated lymphocytes but is quickly transcribed and expressed on T-cells following activation.

This IL-2R is a heterotrimeric protein consisting of three distinct glycopeptide subunits termed IL-2R α (CD25) specific to IL-2R, IL-2R β and IL-2R γ . The α and β chains are involved in binding IL-2 while the signal transduction following IL-2 binding is mediated by the γ -chain along with the β chain. The IL-2R α chain or CD25 is a type 1 transmembrane glycoprotein of 251 amino acids and 55kDa. CD25 can also be found as a soluble form in serum and tissue following enzymatic cleavage from expressing cells and can be identified as a 45KDa protein once shed from the membrane. As the expression and subsequent release of CD25 takes place following cell stimulation the presence of soluble CD25 (sCD25) in circulation is an excellent marker of T-cell activation.

A number of disease states linked to over expression of CD25 have previously been described including, autoimmune diseases, transplant rejection, chronic infection, B-cell neoplasm and various types of leukaemia and other forms of cancer. Because of this definite link between CD25 over expression and disease state many therapies for these conditions have evolved to inhibit this over expression of IL-2R α .

More recently CD25 has become the major marker for distinguishing the CD4+CD25+ subset of T regulatory cells.

2.2. Principle of the method

A capture Antibody highly specific for CD25 has been coated to the wells of the microtiter strip plate provided during manufacture. Binding of CD25 samples and known standards to the capture antibodies and subsequent binding of the biotinylated anti-CD25 secondary antibody to the analyte is completed during the same incubation period. Any excess unbound analyte and secondary antibody is removed. The HRP conjugate solution is then added to every well including the zero wells, following incubation excess conjugate is removed by careful washing. A chromogen substrate is added to the wells resulting in the progressive development of a blue coloured complex with the conjugate. The colour development is then stopped by the addition of acid turning the resultant final product yellow. The intensity of the produced coloured complex is directly proportional to the concentration of CD25 present in the samples and standards. The absorbance of the colour complex is then measured and the generated OD values for each standard are plotted against expected concentration forming a standard curve. This standard curve can then be used to accurately determine the concentration of CD25 in any sample tested.

3. Reagents provided and reconstitution

Reagents (Store@2-8°C)	Quantity 1x48 well kit Cat no. EA102166	Quantity 1x96 well kit Cat no. EA102167	Quantity 2x96 well kit Cat no. EA102168	Reconstitution
96 well microtiter strip plate	1/2	1	2	Ready to use (Pre-coated)
Plastic plate covers	2	2	4	n/a
Standard: 2200pg/ml	1	2	4	Reconstitute as directed on the vial (see reagent preparation, section 8)
Standard Diluent (Buffer)	1 (25ml)	1 (25ml)	1 (25ml)	10x Concentrate, dilute in distilled water (see reagent preparation, section 8)
Control	1	2	4	Reconstitute as directed on the vial (see reagent preparation, section 8)
Biotinylated anti-CD25	1 (0.4ml)	1 (0.4ml)	2 (0.4ml)	Dilute in biotinylated antibody diluent (see reagent preparation, section 8)
Biotinylated Antibody diluent	1 (7ml)	1 (7ml)	1 (13ml)	Ready to use
Streptavidin-HRP	1 (5µl)	2 (5µl)	4 (5µl)	Add 0.5ml of HRP diluent prior to use (see reagent preparation, section 8)
HRP Diluent	1 (23ml)	1 (23ml)	1 (23ml)	Ready to use
Wash Buffer	1 (10ml)	1 (10ml)	2 (10ml)	200x Concentrate dilute in distilled water (see reagent preparation, section 8)
TMB Substrate	1 (11ml)	1 (11ml)	1 (24ml)	Ready to use
H ₂ SO ₄ stop reagent	1 (11ml)	1 (11ml)	2 (11ml)	Ready to use

4. Materials required but not provided

- Microtiter plate reader fitted with appropriate filters (450nm required with optional 620nm reference filter)
- Microplate washer or wash bottle
- 10, 50, 100, 200 and 1,000µl adjustable single channel micropipettes with disposable tips
- 50-300µl multi-channel micropipette with disposable tips
- Multichannel micropipette reagent reservoirs
- Distilled water
- Vortex mixer
- Miscellaneous laboratory plastic and/or glass, if possible sterile

5. Storage Instructions

Store kit reagents between 2 and 8°C. Immediately after use remaining reagents should be returned to cold storage (2-8°C). Expiry of the kit and reagents is stated on box front labels. The expiry of the kit components can only be guaranteed if the components are stored properly, and if, in case of repeated use of one component, the reagent is not contaminated by the first handling.

6. Specimen collection, processing & storage

Cell culture supernatants, human serum, plasma or other biological samples will be suitable for use in the assay. Remove serum from the clot or red cells, respectively, as soon as possible after clotting and separation.

Cell culture supernatants: Remove particulates and aggregates by spinning at approximately 1000 x g for 10 min.

Serum: Use pyrogen/endotoxin free collecting tubes. Serum should be removed rapidly and carefully from the red cells after clotting. Following clotting, centrifuge at approximately 1000 x g for 10 min and remove serum.

Plasma: EDTA, citrate and heparin plasma can be assayed. Spin samples at 1000 x g for 30 min to remove particulates. Harvest plasma.

Storage: If not analyzed shortly after collection, samples should be aliquoted (250-500µl) to avoid repeated freeze-thaw cycles and stored frozen at -70°C. Avoid multiple freeze-thaw cycles of frozen specimens.

Recommendation: Do not thaw by heating at 37°C or 56°C. Thaw at room temperature and make sure that sample is completely thawed and homogeneous before use. When possible avoid use of badly haemolysed or lipemic sera. If large amounts of particles are present these should be removed prior to use by centrifugation or filtration.

7. Safety & precautions for use

- Handling of reagents, serum or plasma specimens should be in accordance with local safety procedures, e.g. CDC/NIH Health manual : " Biosafety in Microbiological and Biomedical Laboratories" 1984.
- Laboratory gloves should be worn at all times.
- Avoid any skin contact with H₂SO₄ and TMB. In case of contact, wash thoroughly with water.
- Do not eat, drink, smoke or apply cosmetics where kit reagents are used.
- Do not pipette by mouth.
- When not in use, kit components should be stored refrigerated or frozen as indicated on vials or bottles labels.
- All reagents should be warmed to room temperature before use. Lyophilized standards should be discarded after use.
- Once the desired number of strips has been removed, immediately reseal the bag to protect the remaining strips from deterioration.
- Cover or cap all reagents when not in use.
- Do not mix or interchange reagents between different lots.
- Do not use reagents beyond the expiration date of the kit.
- Use a clean disposable plastic pipette tip for each reagent, standard, or specimen addition in order to avoid cross contamination, for the dispensing of H₂SO₄ and substrate solution, avoid pipettes with metal parts.
- Use a clean plastic container to prepare the washing solution.
- Thoroughly mix the reagents and samples before use by agitation or swirling.
- All residual washing liquid must be drained from the wells by efficient aspiration or by decantation followed by tapping the plate forcefully on absorbent paper. Never insert absorbent paper directly into the wells.
- The TMB solution is light sensitive. Avoid prolonged exposure to light. Also, avoid contact of the TMB solution with metal to prevent colour development. Warning TMB is toxic avoid direct contact with hands. Dispose off properly.
- If a dark blue colour develops within a few minutes after preparation, this indicates that the TMB solution has been contaminated and must be discarded. Read absorbance's within 1 hour after completion of the assay.
- When pipetting reagents, maintain a consistent order of addition from well-to-well. This will ensure equal incubation times for all wells.
- Follow incubation times described in the assay procedure.
- Dispense the TMB solution within 15 min of the washing of the microtiter plate.

8. Assay Preparation

Bring all reagents to room temperature before use

8.1. Assay Design

Determine the number of microwell strips required to test the desired number of samples plus appropriate number of wells needed for running zeros and standards. Each sample, standard, zero and control should be tested **in duplicate**. Remove sufficient microwell Strips for testing from the pouch immediately prior to use. Return any wells not required for this assay with desiccant to the pouch. Seal tightly and return to 2-8°C storage.

Example plate layout (example shown for a 6 point standard curve)

	Standards / Controls		Sample Wells									
	1	2	3	4	5	6	7	8	9	10	11	12
A	2200	2200										
B	1100	1100										
C	550	550										
D	275	275										
E	137.5	137.5										
F	68.75	68.75										
G	zero	zero										
H	Ctrl	Ctrl										

All remaining empty wells can be used to test samples in duplicate

8.2. Preparation of Wash Buffer

Dilute the (200x) wash buffer concentrate 200 fold with distilled water to give a 1x working solution. Pour entire contents (10 ml) of the Washing Buffer Concentrate into a clean 2,000 ml graduated cylinder. Bring final volume to 2,000 ml with glass-distilled or deionized water. Mix gently to avoid foaming. Transfer to a clean wash bottle and store at 2°-25°C.

8.3. Preparation of Standard Diluent Buffer

Add the contents of the vial (10x concentrate) to 225 ml of distilled water before use.

This Solution can be stored at 2-8°C for up to 1 week.

8.4. Preparation of Standard

Standard vials must be reconstituted with the volume of standard diluent shown on the vial immediately prior to use. This reconstitution gives a stock solution of 2200pg/ml of CD25. Mix the reconstituted standard gently by inversion only. Serial dilutions of the standard are made directly in the assay plate to provide the concentration range from 2200 to 68.75pg/ml. A fresh standard curve should be produced for each new assay.

- Immediately after reconstitution add 200µl of the reconstituted standard to well's A1 and A2, which provides the highest concentration standard at 2200pg/ml.
- Add 100µl of standard diluent to the remaining standard wells B1 and B2 to F1 and F2.
- Transfer 100µl from wells A1 and A2 to B1 and B2. Mix the well contents by repeated aspirations and ejections taking care not to scratch the inner surface of the wells.
- Continue this 1:1 dilution using 100µl from wells B1 and B2 through to wells F1 and F2 providing a serial diluted standard curve ranging from 2200pg/ml to 68.75pg/ml.
- Discard 100µl from the final wells of the standard curve (F1 and F2).

Alternatively these dilutions can be performed in separate clean tubes and immediately transferred into the relevant wells.

8.5. Preparation of Controls

The supplied Controls must be reconstituted with the volume of Standard Diluent indicated on the vial. Reconstitution of the freeze-dried material with the indicated volume, will give a solution at the concentration stated on the vial. Do not store after use.

8.6. Preparation of Samples

Before testing, human serum samples have to be diluted 1:5 in standard buffer diluent.

8.7. Preparation of Biotinylated anti-CD25

It is recommended this reagent is prepared immediately before use. Dilute the biotinylated anti-CD25 with the biotinylated antibody diluent in an appropriate clean glass vial using volumes appropriate to the number of required wells. Please see example volumes below:

Number of wells required	Biotinylated Antibody (µl)	Biotinylated Antibody Diluent (µl)
16	40	1060
24	60	1590
32	80	2120
48	120	3180
96	240	6360

8.8. Preparation of Streptavidin-HRP

It is recommended to centrifuge vial for a few seconds in a microcentrifuge to collect all the volume at the bottom.

Dilute the 5 μ l vial with 0.5ml of HRP diluent **immediately before use**. Do not keep this diluted vial for future experiments. Further dilute the HRP solution to volumes appropriate for the number of required wells in a clean glass vial. Please see example volumes below:

Number of wells required	Streptavidin-HRP (μ l)	Streptavidin-HRP Diluent (ml)
16	30	2
24	45	3
32	60	4
48	75	5
96	150	10

9. Method

We strongly recommend that every vial is mixed thoroughly without foaming prior to use.

Prepare all reagents as shown in section 8.

Note: final preparation of Biotinylated Secondary Antibody (section 8.7) and Streptavidin-HRP (section 8.8) should occur immediately before use.

Assay Step		Details
1.	Addition	Prepare Standard curve as shown in section 8.4 above
2.	Addition	Add 100µl of each, Sample, Standard, Control and zero in duplicate to appropriate number of wells
3.	Addition	Add 50µl of diluted biotinylated anti-CD25 to all wells
4.	Incubation	Cover with a plastic plate cover and incubate at room temperature (18 to 25°C) for 3 hours
5.	Wash	Remove the cover and wash the plate as follows: a) Aspirate the liquid from each well b) Dispense 0.3 ml of 1x washing solution into each well c) Aspirate the contents of each well d) Repeat step b and c another two times
6.	Addition	Add 100µl of Streptavidin-HRP solution into all wells
7.	Incubation	Cover with a plastic plate cover and incubate at room temperature (18 to 25°C) for 30 min
8.	Wash	Repeat wash step 5.
9.	Addition	Add 100µl of ready-to-use TMB Substrate Solution into all wells
10.	Incubation	Incubate in the dark for 10-15 minutes* at room temperature. Avoid direct exposure to light by wrapping the plate in aluminium foil.
11.	Addition	Add 100µl of H₂SO₄: Stop Reagent into all wells
Read the absorbance value of each well (immediately after step 11.) on a spectrophotometer using 450 nm as the primary wavelength and optionally 620 nm as the reference wave length (610 nm to 650 nm is acceptable).		

**Incubation time of the substrate solution is usually determined by the ELISA reader performance. Many ELISA readers only record absorbance up to 2.0 O.D. Therefore the colour development within individual microwells must be observed by the analyst, and the substrate reaction stopped before positive wells are no longer within recordable range.*

10. Data Analysis

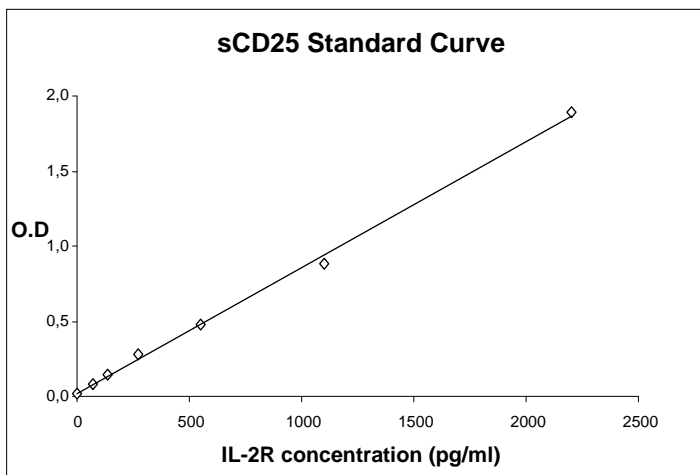
Calculate the average absorbance values for each set of duplicate standards, controls and samples. Ideally duplicates should be within 20% of the mean.

Generate a linear standard curve by plotting the average absorbance of each standard on the vertical axis versus the corresponding CD25 standard concentration on the horizontal axis.

The amount of CD25 in each sample is determined by extrapolating OD values against CD25 standard concentrations using the standard curve.

Example sCD25 Standard curve

Standard	CD25 Conc	OD (450nm) mean	CV (%)
1	2200	1.893	5.4
2	1100	0.883	3.5
3	550	0.473	3.0
4	275	0.258	7.2
5	137.5	0.140	2.2
6	68.25	0.078	1.5
zero	0	0.022	8.9



Note: curve shown above should not be used to determine results. Every laboratory must produce a standard curve for each set of microwell strips assayed.

For samples human serum or plasmas which have been diluted 1:5 according to the protocol, the calculated concentration should be multiplied by the dilution factor (x5).

11. Assay limitations

Do not extrapolate the standard curve beyond the maximum standard curve point. The dose-response is non-linear in this region and good accuracy is difficult to obtain. Concentrated samples above the maximum standard concentration must be diluted with Standard diluent or with your own sample buffer to produce an OD value within the range of the standard curve. Following analysis of such samples always multiply results by the appropriate dilution factor to produce actual final concentration.

The influence of various drugs on end results has not been investigated. Bacterial or fungal contamination and laboratory cross-contamination may also cause irregular results.

Improper or insufficient washing at any stage of the procedure will result in either false positive or false negative results. Completely empty wells before dispensing fresh Washing Buffer, fill with Washing Buffer as indicated for each wash cycle and do not allow wells to sit uncovered or dry for extended periods.

Disposable pipette tips, flasks or glassware are preferred, reusable glassware must be washed and thoroughly rinsed of all detergents before use.

As with most biological assays conditions may vary from assay to assay therefore **a fresh standard curve must be prepared and run for every assay.**

12. Performance Characteristics

12.1. Sensitivity

The sensitivity or minimum detectable dose of CD25 using this OriGeneCD25 ELISA kit was found to be **32.5pg/ml**. This was determined by adding 2 standard deviations to the mean OD obtained when the zero standard was assayed in 6 independent experiments.

12.2. Specificity

The assay recognizes both natural and recombinant human CD25. To define the specificity of this ELISA several proteins were tested for cross reactivity. There was no cross reactivity observed for any protein tested (IL-1 β , IL-2 IL-4, IFN γ , IL-6, IL-6R, TRAIL, IL-7, IL-12 and IL-21).

12.3. Precision

Intra-assay

Reproducibility within the assay will be evaluated in three independent experiments. Each assay will be carried out with 6 replicates (3 duplicates) in 2 human pooled serum, 2 in RPMI and 2 in standard diluent with samples containing different concentrations of CD25. 2 standard curves were run on each plate. **The overall intra-assay coefficient of variation has been calculated to be 4.5%.**

Session	Sample	Mean CD25 pg/ml	SD	CV
Session 1	Sample 1	1389.71	8.20	0.59
	Sample 2	1468.10	31.48	2.14
	Sample 3	1704.83	61.78	3.62
	Sample 4	1268.45	76.20	6.01
	Sample 5	2101.32	80.56	3.83
	Sample 6	1773.03	85.15	4.80
Session 2	Sample 1	1423.01	93.24	6.55
	Sample 2	1536.17	41.66	2.71
	Sample 3	1586.28	32.97	2.08
	Sample 4	944.84	46.03	4.87
	Sample 5	2335.87	117.43	5.03
	Sample 6	1885.74	57.47	3.05
Session 3	Sample 1	1551.81	102.24	6.59
	Sample 2	1578.86	116.29	7.37
	Sample 3	1955.43	111.37	5.70
	Sample 4	952.64	43.74	4.59
	Sample 5	2189.83	116.38	5.31
	Sample 6	1768.18	114.70	6.49

Inter-assay

Assay to assay reproducibility within one laboratory will be evaluated in three independent experiments by two technicians. Each assay will be carried out with 6 replicates (3 duplicates) in 2 human pooled serum, 2 in RPMI and 2 in standard diluent with samples containing different concentrations of CD25. 2 standard curves were run on each plate. **The calculated overall coefficient of variation was 10.3%.**

	Sample 1	Sample2	Sample 3	Sample 4	Sample 5	Sample 6
Mean CD25 pg/ml	1594	1528	1705	1124	2305	1824
SD	184	135	204	171	165	126
CV	11.6	8.8	12.0	15.2	7.2	6.9

12.4. Dilution Parallelism

In two independent experiments two spiked human serum samples with different levels of CD25 were analysed at different serial two fold dilutions (1:4 To 1:16) with two replicates each. Recoveries ranged from 86 to 122% with an overall **mean recovery of 99%.**

12.5. Spike Recovery

The spike recovery was evaluated by spiking 3 concentrations of CD25 in human serum and culture medium in 3 separate experiments. Recoveries ranged from 54 to 106% with an overall **mean recovery of 87%.**

12.6. Stability

Storage Stability

Aliquots of spiked serum samples were stored at -20°C , $2-8^{\circ}\text{C}$, room temperature (RT) and at 37°C and the CD25 level determined after 24h. We observed no loss of activity after storage at 4°C and 37°C . However we did observe a significant loss in activity (21%) following storage at RT. As a result we would advise that samples are stored at either -20°C or 4°C .

Freeze-thaw Stability

Aliquots of spiked serum were stored frozen at -20°C and thawed up to 5 times and the CD25 level was determined. There was no significant loss in activity of CD25 after 5 cycles of freezing and thawing.

12.7. Expected serum values

A panel of 40 human sera and 40 Plasma samples were tested for CD25. See results below:

Sample Matrix	Number of samples evaluated	Range (pg/ml)	Mean (pg/ml)	Standard deviation (pg/ml)
Serum	40	1555-10800	4051	1998
Plasma	40	900-6122	3013	1146

12.8. Standard Calibration

This immunoassay is calibrated against the International Reference Standard NIBSC 97/600. NIBSC 97/600 is quantitated in international units (IU). 1IU corresponding to 57pg OriGeneCD25.

13. Bibliography

Junghans et.al. Biophysical Characterization of a recombinant soluble interleukin 2 receptor. J Biol Chem, 271, pp. 10453-10460. 1996
Morris et.al. Advances in interleukin 2 receptor targeted treatment. Ann Rheum Dis, 59, pp. 109-114. 2000
Nelson B. IL-2, Regulatory T cells and Tolerance. Journal of Immunology, 172, pp. 3983-3988. 2004
Tayal et.al. Cytokines and anti-cytokines as therapeutics- An update. European Journal of Pharmacology, 579, pp. 1-12. 2008
Witkowska A. The role of sIL-2R measurements in Rheumatoid Arthritis and Cancers. Mediators of Inflammation, 3, pp. 121-130. 2005

14. OriGeneHuman CD25 ELISA references

Betjes, M. G. H. et al., Nephrol. Dial. Transplant, 2015: gfv092
Botella-Carretero J. I. et al., Eur. J. Endocrinol., 2005; 153(2): 223 - 230
Dennert, R. et al., Clin. Vaccine Immunol., 2012; 19(8): 1182-1187
Foussat A. et al., Blood, 2001; 98 : 1678 – 168
Magnusson, M. K. et al., United European Gastroenterology Journal, 2013: 2050640613502962
Malmstrom, P.-U. et al., Clin. Cancer Res., 2010; 16(12): 3279-3287
Marcellis, R.G.J. et al., Eur. Respir. J., 2011; 38: 628 – 634
Mostard, R. L. et al., BMC Pulm Med, 2012; 12(1): 57
Verwoerd, A. et al., Eur. Respir. J., 2015: 09031936.00005315
Wijnen, P. A. et al., Eur. Respir. J., 2014 ; 43(6): 1730-1739

15. Assay Summary

Total procedure length: 3h45mn

Add 100 μ l of sample and diluted standard/controls and 50 μ l Biotinylated anti-CD25



Incubate 3 hours at room temperature



Wash three times



Add 100 μ l of Streptavidin-HRP



Incubate 30min at room temperature



Wash three times



Add 100 μ l of ready-to-use TMB
Protect from light. Let the color develop for 10-15 mn.



Add 100 μ l H₂SO₄



Read Absorbance at 450 nm

TECHNICAL CONSULTATION

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For Research Use Only
Not for use in diagnostic procedures

16. International Summaries

16.1. French

PREPARATION DES REACTIFS : RESUME

1. Tampon de Lavage (Washing Buffer) Ajouter 10 ml de **Tampon de Lavage concentré** (*Washing Buffer Concentrate*) 200 fois (200X) à 1990 ml d'eau distillée

2 Tampon de Dilution du Standard (*Standard Diluent Buffer*) Ajouter 25 ml de **Tampon de Dilution du Standard concentré** 10 fois (*Standard Diluent Buffer Concentrate 10X*) à 225 ml d'eau distillée

3. Standard CD25 (*CD25 Standard*) Reconstituer le **Standard CD25** en ajoutant la quantité indiquée sur le flacon de Tampon de Dilution du Standard

4. Contrôles (*CD25 Controls*) Reconstituer les **Contrôles** (*Controls*) en ajoutant la quantité indiquée sur le flacon de Tampon de Dilution du Standard

5. Anti-CD25 Biotinylé (<i>Biotinylated anti-sCD25</i>)	Nombre de barrettes	Anti-CD25 Biotinylé Concentré (µl)	Diluent de l'Anticorps Biotinylé (µl)
	2	40	1060
	3	60	1590
	4	80	2120
	6	120	3180
	12	240	6360

6. Streptavidin-HRP (<i>Streptavidin-HRP</i>)	Nombre de barrettes	Streptavidin-HRP pré- diluée (µl)	Diluent HRP (ml)
	2	30	2
	3	45	3
	4	60	4
	6	75	5
	12	150	10

RESUME DU PROTOCOLE OPERATOIRE: durée totale : 3h45mn

1. Ajouter 100 µl de **Tampon de Dilution du Standard** (*Standard Diluent Buffer*), en duplicat, dans les puits Standards (B1 à F2).
2. Ajouter à la pipette 200 µl de **Standard CD25** (*CD25 Standard*) reconstitué dans les puits A1 et A2 puis réaliser des dilutions du Standard allant de 2200 à 68.75 pg/ml en transférant 100 µl d'un puits à l'autre. Jeter les 100 µl des derniers puits (F1 et F2).
3. Ajouter 100 µl de **Tampon de Dilution du Standard** (*Standard Diluent Buffer*) en duplicat dans les puits "blancs".
4. Ajouter 100 µl d'**échantillon** (*Sample*), en duplicat, dans les puits désignés et 100 µl de contrôle (*CD25 control*) en duplicat dans les puits contrôles.
5. Préparer l'**anticorps anti-CD25 Biotinylé** (*Biotinylated anti CD25*).
6. Ajouter 50 µl d'anticorps **anti-CD25 Biotinylé dilué** (*diluted biotinylated anti CD25*) dans tous les puits.
7. Couvrir les barrettes de puits et incuber pendant 3 heures à température ambiante (18-25°C).
8. Vider et laver les puits 3 fois avec le **Tampon de Lavage** (*Washing Buffer*).
9. Préparer la Streptavidin-HRP.
10. Ajouter 100 µl de **Streptavidin-HRP diluée** (*diluted HRP-Streptavidin*) dans tous les puits.
11. Couvrir les puits et incuber pendant 30 minutes à température ambiante (18-25°C).
12. Vider et laver les puits 3 fois avec le **Tampon de Lavage** (*Washing Buffer*).
13. Ajouter 100 µl de solution de TMB (*TMB solution*) prête à l'emploi dans tous les puits y compris les "blancs".
14. Incuber pendant environ 10-15 minutes à température ambiante (18-25°C) à l'obscurité.
15. Ajouter 100 µl d' H_2SO_4 : **Solution Stop** (H_2SO_4 : *Stop Solution*) dans tous les puits y compris les "blancs".
16. Mesurer l'absorbance (Densité Optique = D.O.) à la longueur d'onde 450 nm et optionnellement à 620 nm (entre 610 et 650 nm) comme longueur d'onde de référence.

Remarque: Les échantillons présentant une valeur de D.O. excédant la gamme de la courbe Standard peuvent résulter à des taux de CD25 incorrects. C'est pourquoi, il est recommandé de diluer de tels échantillons avec le Tampon de Dilution du Standard (*Standard Diluent Buffer*) afin de quantifier précisément le véritable taux de CD25.

16.2. Spanish

PREPARACIÓN DE LOS PRODUCTOS

1. Tampón de Lavado (Washing Buffer) Añadir **Tampón de Lavado Concentrado** 200 X (10 ml) (*Washing Buffer Concentrate*) a 1990 ml de agua destilada.

2. Tampón diluyente del estándar (*Standard Diluent Buffer*) Añadir **Tampón Diluyente del Estándar Concentrado** 10 X (25 ml) (*Standard diluent buffer concentrate 10X*) a 225 ml de agua destilada.

3. Estándar CD25 (*CD25 Standard*) Reconstituir el **Estándar CD25** añadiendo el Diluyente del Estándar, como indica la etiqueta del vial.

4. Controles CD25 (*CD25 Control*) Reconstituir los **controles** añadiendo el Diluyente del Estándar, como indica la etiqueta del vial.

5. Anti-CD25 biotinilado (<i>Biotinylated anti-CD25</i>)	Número de tiras	Anticuerpo biotinilado concentrado (µl)	Diluyente del anticuerpo biotinilado (µl)
	2	40	1,060
	3	60	1,590
	4	80	2,120
	6	120	3,180
	12	240	6,360

6. Estreptavidina-HRP (<i>Streptavidin-HRP</i>)	Número de tiras	Estreptavidina-HRP prediluida (µl)	Diluyente de HRP (ml)
	2	30	2
	3	45	3
	4	60	4
	6	75	5
	12	150	10

RESUMEN DEL PROTOCOLO. El procedimiento total tiene una duración de 3h45min.

1. Añadir 100 µl del **Tampón Diluyente del Estándar** (*Standard Diluent Buffer Concentrate 10X*), por duplicado, a los pocillos designados para el estándar (B1 to F2).
2. Pipetear 200 µl del **Estándar CD25** (*CD25 Standard*) reconstituido en los pocillos A1 y A2 y hacer diluciones seriadas del estándar con el rango de concentraciones de 2200 al 68.75 pg/ml, transfiriendo 100 µl de un pocillo al siguiente. Descartar 100 µl de los últimos pocillos.
3. Añadir 100 µl del **Tampón Diluyente del Estándar**, por duplicado, a los pocillos que van a ser el “blanco”.
4. Añadir 100 µl de las muestras, por duplicado, a los pocillos designados para ello, y 100 µl del Control reconstituido (*CD25 Control*), por duplicado, a los pocillos designados como “control”.
5. Preparar el anticuerpo **Anti-CD25 Biotinilado** (*Biotinylated anti-sCD25*).
6. Añadir 50 µl del **anti-CD25 Biotinilado** y diluido, a todos los pocillos.
7. Cubrir la placa e incubar durante 3 horas a temperatura ambiente (18-25°C).
8. Vaciar y lavar la placa 3 veces con **Tampón de Lavado** (*Washing Buffer*).
9. Preparar la **Estreptavidina-HRP** (*Streptavidin-HRP*).
10. Añadir 100 µl de **Estreptavidina-HRP** diluida a todos los pocillos.
11. Cubrir la placa e incubar 30 minutos a temperatura ambiente (18-25°C).
12. Vaciar y lavar la placa 3 veces con **Tampón de Lavado**.
13. Añadir 100 µl de solución **TMB preparado para utilizar** (*TMB Substrate*), a todos los pocillos, incluidos los pocillos con “blancos”.
14. Incubar la placa durante 10-15 minutos a temperatura ambiente (18-25°C) y en oscuridad.
15. Añadir 100 µl de H₂SO₄: **Solución de Parada** (*H₂SO₄ Stop Solution*), a todos los pocillos, incluidos los pocillos con los “blancos”.
16. Medir la intensidad de color (densidad óptica) a 450 nm y a 620 nm como longitud de onda de referencia (de 610 nm a 650 nm sería aceptable).

Nota: El cálculo de concentraciones de muestras con densidad óptica que supere el rango de la curva estándar, resultaría incorrecto, dando niveles de CD25 más bajos de lo real. Estas muestras, requerirían ser diluidas con el Tampón de Dilución de Estándar, para poder precisar la cantidad real de CD25.