



Human IL-17F ELISA Kit

Instructions for use

Catalogue numbers: 1x48 tests: EA101271
 1x96 tests: EA101272
 2x96 tests: EA101273

For research use only

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Human IL-17F ELISA KIT

1. Intended use

The OriGeneIL-17F ELISA kit is a solid phase sandwich ELISA for the *in vitro* qualitative and quantitative determination of IL-17F in supernatants, buffered solutions or serum and plasma samples and other body fluids. This assay will recognise both natural and recombinant human IL-17F.

This kit has been configured for research use only.

2. Introduction

2.1. Summary

IL-17F, discovered in 2001, is a member of the IL-17 family of cytokines produced by activated T-cells CD4+ and monocytes. IL-17F has been considered as an inflammatory cytokine since it induces many proinflammatory cytokines and chemokines. IL-17F shares strongest homology to IL-17A (sharing about 50% amino acid). The IL-17A and IL-17F genes are localized in the same chromosome region. An IL-17A/F heterodimer biologically active was found to be expressed in Th17 cells together with IL-17A and IL-17F homodimers.

2.2. Principle of the method

A capture Antibody highly specific for IL-17F has been coated to the wells of the microtitre strip plate provided during manufacture. Binding of IL-17F in samples and known standards to the capture antibodies is completed and then any excess unbound analyte is removed. During the next incubation period the binding of the biotinylated anti-IL-17F secondary antibody to the analyte occurs. Any excess unbound secondary antibody is then removed. The HRP conjugate solution is then added to every well including the zero wells, following incubation excess conjugate is removed by careful washing. A chromogen substrate is added to the wells resulting in the progressive development of a blue coloured complex with the conjugate. The colour development is then stopped by the addition of acid turning the resultant final product yellow. The intensity of the produced coloured complex is directly proportional to the concentration of IL-17F present in the samples and standards. The absorbance of the colour complex is then measured and the generated OD values for each standard are plotted against expected concentration forming a standard curve. This standard curve can then be used to accurately determine the concentration of IL-17F in any sample tested.

3. Reagents provided and reconstitution

Reagents (Store @ 2-8°C)	Quantity 1x48 well kit Cat no. EA101271	Quantity 1x96 well kit Cat no. EA101272	Quantity 2x96 well kit Cat no. EA101273	Reconstitution
96 well microtiter strip plate	1/2	1	2	Ready to use (Pre-coated)
Plastic plate covers	2	2	4	n/a
Standard: 500 pg/ml	1 vial	2 vials	4 vials	Reconstitute as directed on the vial (see Assay preparation, section 8)
Standard Diluent (Buffer)	1 vial (25ml)	1 vial (25ml)	2 vials (25ml)	Ready to use
Biotinylated anti- IL-17F	1 vial (0.4ml)	1 vial (0.4ml)	2 vials (0.4ml)	Dilute in Biotinylated Antibody Diluent (see Assay preparation, section 8)
Biotinylated Antibody Diluent	1 vial (7ml)	1 vial (7ml)	1 vial (13ml)	Ready to use
Streptavidin-HRP	1 vial (5µl)	2 vials (5µl)	4 vials (5µl)	Add 5µl of HRP diluent prior to use (see Assay preparation, section 8)
HRP Diluent	1 vial (23ml)	1 vial (23ml)	1 vial (23ml)	Ready to use
Wash Buffer	1 vial (10ml)	1 vial (10ml)	2 vials (10ml)	(10ml) 200x Concentrate dilute in distilled water (see Assay preparation, section 8)
TMB Substrate	1 vial (11ml)	1 vial (11ml)	1 vial (24ml)	Ready to use
H ₂ SO ₄ stop reagent	1 vial (11ml)	1 vial (11ml)	2 vials (11ml)	Ready to use

4. Materials required but not provided

- Microtitre plate reader fitted with appropriate filters (450nm required with optional 620nm reference filter)
- Microplate washer or wash bottle
- 10, 50, 100, 200 and 1,000µl adjustable single channel micropipettes with disposable tips
- 50-300µl multi-channel micropipette with disposable tips
- Multichannel micropipette reagent reservoirs
- Distilled water
- Vortex mixer
- Miscellaneous laboratory plastic and/or glass, if possible sterile

5. Storage Instructions

Store kit reagents between 2 and 8°C. Immediately after use remaining reagents should be returned to cold storage (2-8°C). Expiry of the kit and reagents is stated on box front labels. The expiry of the kit components can only be guaranteed if the components are stored properly, and if, in case of repeated use of one component, the reagent is not contaminated by the first handling.

6. Specimen collection, processing & storage

Cell culture supernatants, human serum, plasma or other biological samples will be suitable for use in the assay. Remove serum from the clot or red cells, respectively, as soon as possible after clotting and separation.

Cell culture supernatants: Remove particulates and aggregates by spinning at approximately 1000 x g for 10 min.

Serum: Use pyrogen/endotoxin free collecting tubes. Serum should be removed rapidly and carefully from the red cells after clotting. Following clotting, centrifuge at approximately 1000 x g for 10 min and remove serum.

Plasma: EDTA, citrate and heparin plasma can be assayed. Spin samples at 1000 x g for 30 min to remove particulates. Harvest plasma.

Storage: If not analyzed shortly after collection, samples should be aliquoted (250-500µl) to avoid repeated freeze-thaw cycles and stored frozen at -70°C . Avoid multiple freeze-thaw cycles of frozen specimens.

Recommendation: Do not thaw by heating at 37°C or 56°C . Thaw at room temperature and make sure that sample is completely thawed and homogeneous before use. When possible avoid use of badly haemolysed or lipemic sera. If large amounts of particles are present these should be removed prior to use by centrifugation or filtration.

7. Safety & precautions for use

- Handling of reagents, serum or plasma specimens should be in accordance with local safety procedures , e.g.CDC/NIH Health manual : " Biosafety in Microbiological and Biomedical Laboratories" 1984
- Avoid any skin contact with H₂SO₄ and TMB. In case of contact, wash thoroughly with water
- Do not eat, drink, smoke or apply cosmetics where kit reagents are used
- Do not pipette by mouth
- When not in use, kit components should be stored refrigerated or frozen as indicated on vials or bottles labels
- All reagents should be warmed to room temperature before use. Lyophilized standards should be discarded after use
- Once the desired number of strips has been removed, immediately reseal the bag to protect the remaining strips from deterioration
- Cover or cap all reagents when not in use
- Do not mix or interchange reagents between different lots
- Do not use reagents beyond the expiration date of the kit
- Use a clean disposable plastic pipette tip for each reagent, standard, or specimen addition in order to avoid cross contamination, for the dispensing of H₂SO₄ and substrate solution, avoid pipettes with metal parts
- Use a clean plastic container to prepare the washing solution
- Thoroughly mix the reagents and samples before use by agitation or swirling
- All residual washing liquid must be drained from the wells by efficient aspiration or by decantation followed by tapping the plate forcefully on absorbent paper. Never insert absorbent paper directly into the wells
- The TMB solution is light sensitive. Avoid prolonged exposure to light. Also, avoid contact of the TMB solution with metal to prevent colour development. Warning TMB is toxic avoid direct contact with hands. Dispose off properly
- If a dark blue colour develops within a few minutes after preparation, this indicates that the TMB solution has been contaminated and must be discarded. Read absorbance's within 1 hour after completion of the assay
- When pipetting reagents, maintain a consistent order of addition from well-to-well. This will ensure equal incubation times for all wells
- Follow incubation times described in the assay procedure
- Dispense the TMB solution within 15 min of the washing of the microtitre plate

8. Assay Preparation

Bring all reagents to room temperature before use

8.1. Assay Design

Determine the number of microwell strips required to test the desired number of samples plus appropriate number of wells needed for running zeros and standards. Each sample, standard and zero should be tested **in duplicate**. Remove sufficient Microwell Strips for testing from the aluminium pouch immediately prior to use. Return any wells not required for this assay with desiccant to the pouch. Seal tightly and return to 2-8°C storage.

Example plate layout(example shown for a 6 point standard curve)

	Standards		Sample Wells									
	1	2	3	4	5	6	7	8	9	10	11	12
A	500	500										
B	250	250										
C	125	125										
D	62.5	62.5										
E	31.25	31.25										
F	15.6	15.6										
G	zero	zero										
H												

All remaining empty wells can be used to test samples in duplicate

8.2. Preparation of Wash Buffer

Dilute the (200x) wash buffer concentrate 200 fold with distilled water to give a 1x working solution. Pour entire contents (10 ml) of the Washing Buffer Concentrate into a clean 2,000 ml graduated cylinder. Bring final volume to 2,000 ml with glass-distilled or deionized water. Mix gently to avoid foaming. Transfer to a clean wash bottle and store at 2°-8°C for up to 1 week.

8.3. Preparation of Standard

Standard vials must be reconstituted with the volume of standard diluent shown on the vial immediately prior to use. This reconstitution gives a stock solution of 500pg/ml of IL-17F. **Mix the reconstituted standard gently by inversion only.** Serial dilutions of the standard are made directly in the assay plate to provide the concentration range from 500 to 15.6pg/ml. A fresh standard curve should be produced for each new assay.

- Immediately after reconstitution add 200µl of the reconstituted standard to well's A1 and A2, which provides the highest concentration standard at 500pg/ml
- Add 100µl of appropriate standard diluent to the remaining standard wells B1 and B2 to F1 and F2
- Transfer 100µl from wells A1 and A2 to B1 and B2. Mix the well contents by repeated aspirations and ejections taking care not to scratch the inner surface of the wells
- Continue this 1:1 dilution using 100µl from wells B1 and B2 through to wells F1 and F2 providing a serial diluted standard curve ranging from 500pg/ml to 15.6pg/ml
- Discard 100µl from the final wells of the standard curve (F1 and F2)

Alternatively these dilutions can be performed in separate clean tubes and immediately transferred directly into the relevant wells.

8.4. Preparation of Biotinylated anti-IL-17F

It is recommended this reagent is prepared immediately before use. Dilute the biotinylated anti-IL-17F with the biotinylated antibody diluent in an appropriate clean glass vial using volumes appropriate to the number of required wells. Please see example volumes below:

Number of wells required	Biotinylated Antibody (μ l)	Biotinylated Antibody Diluent (μ l)
16	40	1060
24	60	1590
32	80	2120
48	120	3180
96	240	6360

8.5. Preparation of Streptavidin-HRP

It is recommended to centrifuge vial for a few seconds in a microcentrifuge to collect all the volume at the bottom.

Dilute the 5 μ l vial with 0.5ml of HRP diluent **immediately before use**. Do-not keep this diluted vial for future experiments. Further dilute the HRP solution to volumes appropriate for the number of required wells in a clean glass vial. Please see example volumes below:

Number of wells required	Streptavidin-HRP (μ l)	Streptavidin-HRP Diluent (ml)
16	30	2
24	45	3
32	60	4
48	75	5
96	150	10

9. Method

We strongly recommend that every vial is mixed thoroughly without foaming prior to use except the standard vial which must be mixed gently by inversion only.

Prepare all reagents as shown in section 8.

Note: Final preparation of Biotinylated anti-IL-17F (section 8.5) and Streptavidin-HRP (section 8.6) should occur immediately before use.

Assay Step		Details
1.	Addition	Prepare Standard curve as shown in section 8.4
2.	Addition	Add 100µl of standard diluent to zero wells and 50µl to sample wells if serum or plasma are tested
3.	Addition	Add 100µl of each Sample in duplicate to appropriate number of wells if using cell culture supernatant or buffered solutions or 50µl if using human serum or plasma
4.	Incubation	Cover with a plastic plate cover and incubate at room temperature (18 to 25°C) for 2 hour(s)
5.	Wash	Remove the cover and wash the plate as follows: a) Aspirate the liquid from each well b) Dispense 0.3 ml of 1x washing solution into each well c) Aspirate the contents of each well d) Repeat step b and c another two times
6.	Addition	Add 50µl of diluted biotinylated anti-IL-17F to all wells
7.	Incubation	Cover with a plastic plate cover and incubate at room temperature (18 to 25°C) for 1 hour(s)
8.	Wash	Repeat wash step 4.
9.	Addition	Add 100µl of Streptavidin-HRP solution into all wells
10.	Incubation	Cover with a plastic plate cover and incubate at room temperature (18 to 25°C) for 30 min
11.	Wash	Repeat wash step 4.
12.	Addition	Add 100µl of ready-to-use TMB Substrate Solution into all wells
13.	Incubation	Incubate in the dark for 10-20 minutes* at room temperature. Avoid direct exposure to light by wrapping the plate in aluminium foil.
14.	Addition	Add 100µl of H₂SO₄:Stop Reagent into all wells
Read the absorbance value of each well (immediately after step 14.) on a spectrophotometer using 450 nm as the primary wavelength and optionally 620 nm as the reference wave length (610 nm to 650 nm is acceptable).		

*Incubation time of the substrate solution is usually determined by the ELISA reader performance. Many ELISA readers only record absorbance up to 2.0 O.D. Therefore the colour development within individual microwells must be observed by the analyst, and the substrate reaction stopped before positive wells are no longer within recordable range

10. Data Analysis

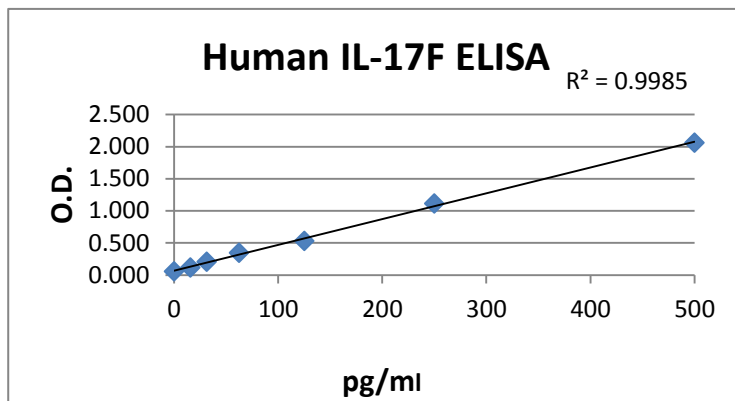
Calculate the average absorbance values for each set of duplicate standards and samples. Ideally duplicates should be within 20% of the mean.

Generate a linear standard curve by plotting the average absorbance of each standard on the vertical axis versus the corresponding IL-17F standard concentration on the horizontal axis.

The amount of IL-17F in each sample is determined by extrapolating OD values against IL-17F standard concentrations using the standard curve.

Example IL-17F Standard curve

Standard	IL-17F Conc	OD (450nm) mean	CV (%)
1	500.0	2.061	2.9
2	250.0	1.114	2.9
3	125.0	0.530	9.7
4	62.5	0.343	5.4
5	31.25	0.203	0.9
6	15.6	0.117	4.8
zero	0.0	0.054	4.8



Note; curve shown above should not be used to determine results. Every laboratory must produce a standard curve for each set of microwell strips assayed.

If serum or plasma samples have been assayed following the instructions of the protocol, the concentration read from the standard curve must be multiplied by the dilution factor (x2) (50µl sample + 50µl standard diluent).

11. Assay limitations

Do not extrapolate the standard curve beyond the maximum standard curve point. The dose-response is non-linear in this region and good accuracy is difficult to obtain. Concentrated samples above the maximum standard concentration must be diluted with Standard diluent or with your own sample buffer to produce an OD value within the range of the standard curve. Following analysis of such samples always multiply results by the appropriate dilution factor to produce actual final concentration.

The influence of various drugs on end results has not been investigated. Bacterial or fungal contamination and laboratory cross-contamination may also cause irregular results.

Improper or insufficient washing at any stage of the procedure will result in either false positive or false negative results. Completely empty wells before dispensing fresh Washing Buffer, fill with Washing Buffer as indicated for each wash cycle and do not allow wells to sit uncovered or dry for extended periods.

Disposable pipette tips, flasks or glassware are preferred, reusable glassware must be washed and thoroughly rinsed of all detergents before use.

As with most biological assays conditions may vary from assay to assay therefore **afresh standard curve must be prepared and run for every assay.**

12. Performance Characteristics

12.1. Sensitivity

The sensitivity, minimum detectable dose of IL-17F using this OriGenIL-17F ELISA kit was found to be **6.0pg/ml**. This was determined by adding 3 standard deviations to the mean OD obtained when the zero standard was assayed 40 times.

12.2. Specificity

The assay recognizes natural human IL-17F. To define specificity of this ELISA, several proteins were tested for cross reactivity. There was no cross reactivity observed for these proteins tested IL-17A, IL-17B, IL-17D, IL17E, IL-5, IL-23, Perforin. The kit shows cross reactivity with the human IL17A/F heterodimer.

12.3. Precision

The reproducibility was evaluated in three experiments with 6 replicates of 3 different samples containing different levels of IL-17F. The data below shows the mean concentrations and the CV values for each sample. **The overall intra-assay CV was 7.1% and inter-assay was 7.7%.**

Intra-Assay					Inter-Assay				
Sample	n	Mean (pg/mL)	SD	CV%	Sample	n	Mean (pg/mL)	SD	CV%
A	9	72	8	11.2	A	9	72	8.6	11.9
B	9	76	5.8	7.7	B	9	76	5.9	7.7
C	9	112	2.8	2.5	C	9	112	3.8	3.4

12.4. Dilution Parallelism

Two spiked human serum with different levels of recombinant IL-17F were analysed at three serial two fold dilutions (1:2-1:8) with two replicates each. Recoveries ranged from 91% to 111% with an overall **mean recovery of 100%**.

12.5. Spike Recovery

The spike recovery was evaluated by spiking three concentrations of recombinant IL-17F in human serum in two experiments. Recoveries ranged from 68% to 110% with an overall **mean recovery of 89%**.

12.6. Stability

Storage Stability

Aliquots of spiked serum samples were stored at -20°C , $2-8^{\circ}\text{C}$, room temperature (RT) and at 37°C and the IL-17F level determined after 24h. We observed a loss of 30% after storage at $2-8^{\circ}\text{C}$ and 40% after storage at RT and 37°C .

Freeze-thaw Stability

Aliquots of spiked serum were stored frozen at -20°C and thawed up to 5 times and IL-17F level was determined. We observed a loss of up to 42% of IL-17F reactivity during storage.

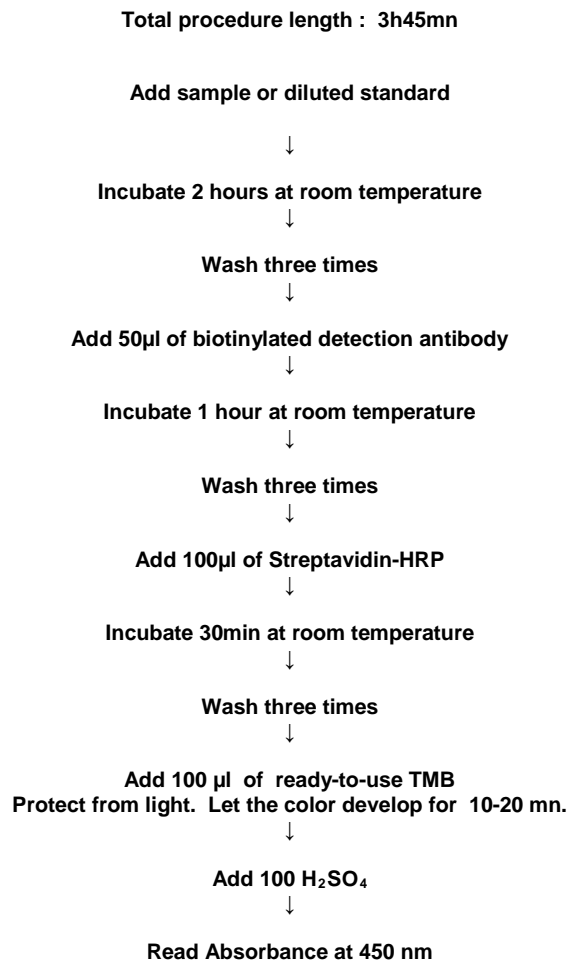
12.7. Expected serum values

A panel of 20 apparently healthy blood donors was tested for IL-17F. All were below the detection level <6 pg/ml.

13. Bibliography

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14. Assay Summary



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