

## Product Information

### Rubella IgG ELISA kit

Catalog Number: EA100957

Storage Temperature: 2 – 8°C

## Instruction for Use

### Intended Use

The Rubella IgG ELISA Kit is intended for the detection of IgG antibody to Rubella in human serum or plasma.

### Background

Rubella is usually a mild disease with infrequent complication. In unvaccinated populations, rubella is primarily a childhood disease. Where children are well-immunized, adolescent and adult infections become more evident. Rubella is spread by direct contact with nasal or throat secretions of infected individuals. Symptoms may include a rash, slight fever, joint aches, headache, discomfort, runny nose and reddened eyes. The incubation period for rubella is 12-23 days; in most cases, symptoms appear within 16-18 days. If contracted during the first trimester of pregnancy, Rubella infection can lead to congenital rubella syndrome (CRS). Infection of a pregnant woman may result in a miscarriage, stillbirth or the birth of an infant with abnormalities, which may include deafness, cataracts, heart defects, liver and spleen damage and mental retardation. CRS occurs among at least 25 percent of infants born to women who have had rubella during the first trimester of pregnancy. The presence of IgG antibody to rubella virus is indicative of vaccination or previous exposure. In individuals with acute rubella infection, four-fold or greater increase in IgG antibody level is indicative of recent infection. Rubella IgM antibodies are detected by ELISA in 100% of patients between days 11-25 after onset of the exanthema, in 60-80% of individuals at days 15-25 after vaccination and in 90-97% of infants with congenital rubella between 2 weeks and 3 months after birth. Rubella IgM antibody often persists for 20-30 days after acute infection or vaccination.

### Principle of the Test

Diluted patient serum is added to wells coated with purified antigen. IgG specific antibody, if present, binds to the antigen. All unbound materials are washed away and the enzyme conjugate is added to bind to the antibody-antigen complex, if present. Excess enzyme conjugate is washed off and substrate is added. The plate is incubated to allow the hydrolysis of the substrate by the enzyme. The intensity of the color generated is proportional to the amount of IgG specific antibody in the sample.

### Components

<b>MATERIALS PROVIDED</b>	<b>96 Tests</b>
1. Microwells coated with Rubella antigen	12x8x1
2. Sample Diluent: 1 bottle (ready to use)	22 ml
3. Calibrator: 1 Vial (ready to use)	1ml

4. Positive Control: 1 vial (ready to use)	1ml
5. Negative Control: 1 vial (ready to use)	1ml
6. Enzyme conjugate: 1 bottle (ready to use)	12ml
7. TMB Substrate: 1 bottle (ready to use)	12ml
8. Stop Solution: 1 bottle (ready to use)	12ml
9. Wash concentrate 20X: 1 bottle	25ml

### Materials and Equipment Required but Not Provided

1. Distilled or deionized water
2. Precision pipettes
3. Disposable pipette tips
4. ELISA reader capable of reading absorbance at 450nm
5. Absorbance paper or paper towel

### Disclaimer

This product is for research use only and not intended for diagnostic procedures.

### Specimen Collection and Preparation

1. Collect blood specimens and separate the serum.
2. Specimens may be refrigerated at 2–8°C for up to seven days or frozen for up to six months. Avoid repetitive freezing and thawing.

### Reagent Preparation

1. Prepare 1X Wash buffer by adding Wash Concentrate (25 ml, 20X) to 475 ml of distilled or deionized water. Store at room temperature (18-26°C).

### Assay Procedure

- Before proceeding with the assay, bring all reagents, serum references and controls to room temperature (18-26°C). Gently mix all reagents before use
  - The components in this kit are intended for use as an integral unit. The components of different lots should not be mixed
  - It is recommended that standards, control and serum samples be run in duplicate
  - Do not use sodium azide as preservative. Sodium azide inhibits HRP enzyme activities
1. Place the desired number of coated strips into the holder.
  2. Negative control, positive control, and calibrator are ready to use. Prepare 1:21 dilution of test samples, by adding 10 µl of the sample to 200 µl of sample diluent. Mix well.
  3. Dispense 100 µl of diluted sera, calibrator and controls into the appropriate wells. For the reagent blank, dispense 100 µl sample diluent in 1A well position. Tap the holder to remove air bubbles from the liquid and mix well. Incubate for 20 minutes at room temperature.
  4. Remove liquid from all wells. Wash wells three times with 300 µl of 1X wash buffer. Blot on absorbance paper or paper towel.
  5. Dispense 100 µl of enzyme conjugate to each well and incubate for 20 minutes at room temperature.
  6. Remove enzyme conjugate from all wells. Wash wells three times with 300 µl of 1X wash buffer. Blot on absorbance paper or paper towel.

7. Dispense 100  $\mu$ l of TMB substrate and incubate for 10 minutes at room temperature. Add 100  $\mu$ l of stop solution.
8. Read O.D. at 450 nm using ELISA reader within 15 min. A dual wavelength is recommended with reference filter of 600-650 nm

### Calculation of Results

1. Check Calibrator Factor (CF) value on the calibrator bottle. This value might vary from lot to lot. Make sure you check the value on every kit.
2. Calculate the cut-off value: Calibrator OD x Calibrator Factor (CF).
3. Calculate the Ab (Antibody) Index of each determination by dividing the O.D. value of each sample by cut-off value.

### Example of a Standard Curve

Calibrator mean OD = 0.8

Calibrator Factor (CF) = 0.5

Cut-off Value =  $0.8 \times 0.5 = 0.400$

Positive control O.D. = 1.2

Ab Index =  $1.2 / 0.4 = 3$

Patient sample O.D. = 1.6

Ab Index =  $1.6 / 0.4 = 4.0$

### Quality Control

The test run may be considered valid provided the following criteria are met:

1. If the O.D. of the Calibrator should be greater than 0.250.
2. The Ab index for Negative control should be less than 0.9.
3. The Ab index for Positive control should be greater than 1.2.

### Interpretation

The following is intended as a guide to interpretation of Rubella IgG test results; each laboratory is encouraged to establish its own criteria for test interpretation based on sample populations encountered.

#### • Antibody Index Interpretation

- <0.9 No detectable antibody to Rubella IgG by ELISA
- 0.9-1.1 Borderline positive. Follow-up testing is recommended if clinically indicated.
- >1.1 Detectable antibody to Rubella IgG by ELISA

### References

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2. Matter L; Germann D; Bally F; Schopfer K. Age-stratified seroprevalence of Rubella, mumps and rubella (MMR) virus infections in Switzerland after the introduction of MMR mass vaccination. Eur J Epidemiol 1997;13(1):61-6.
3. M'uhlebach-Sponer M; Zbinden R; da Silva VA; Gnehm HE. Intrathecal rubella antibodies in an adolescent with Guillain-Barré syndrome after mumps-Rubella-rubella vaccination [letter]. Eur J Pediatr 1995; 154(2):166.

4. Johnson CE; Kumar ML; Whitwell JK; Staehle BO; Rome LP; Dinakar C; Hurni W; Nalin DR. Antibody persistence after primary Rubella-mumps-rubella vaccine and response to a second dose given at four to six vs. eleven to thirteen years. *Pediatr Infect Dis J* 1996;15(8):687-92.
5. Matter L; Kogelschatz K; Germann D. Serum levels of rubella virus antibodies indicating immunity: response to vaccination of subjects with low or undetectable antibody concentrations. *J Infect Dis* 1997; 175(4):749-55.
6. Bos P; Steele D; Alexander J. Prevalence of antibodies to rubella, herpes simplex 2 and cytomegalovirus in pregnant women and in neonates at Ga-Rankuwa. *Cent Afr J Med* 1995;41(1):14-7.

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