

## Product Information

### Thyroglobulin Antibody (TG) ELISA kit

Catalog Number: EA100925

Storage Temperature: 2 – 8°C

## Instruction for Use

### Intended Use

The Thyroglobulin (TG) Ab ELISA Kit is intended for the detection of IgG antibody to Thyroglobulin (TG) in human serum or plasma.

### Background

Thyroglobulin is a water soluble glycoprotein that is involved in the storage and synthesis of thyroid hormones. The thyroid microsomal antigen has been shown to be the enzyme thyroid peroxidase (TPO). Antibodies to thyroglobulin and or microsomal antigen are present in most patients with goitrous thyroiditis (Hashimoto disease), atrophic thyroiditis and about 70-90% of Graves disease. Antibodies are also found in about half of the patients with primary hypothyroidism and thyrotoxicosis, and 10-20% of patients with simple goiters and thyroid tumors. There is also a relationship between thyroid antibodies and diabetes mellitus. Thyroid autoantibodies are present in about 6-7% of normals and their incidence increases with age. Classically, autoantibodies to thyroid antigens are detected by precipitation reactions, hemagglutination and by immunofluorescence. However the tests are subjective and lack high sensitivity. Enzyme-Linked Immunosorbent Assays (ELISAs) combine greater sensitivity, objective reading and ease of use. ELISAs have been developed and validated for detecting autoantibodies to thyroid antigens.

### Principle of the Test

Diluted patient serum is added to wells coated with purified antigen. IgG specific antibody, if present, binds to the antigen. All unbound materials are washed away and the enzyme conjugate is added to bind to the antibody-antigen complex, if present. Excess enzyme conjugate is washed off and substrate is added. The plate is incubated to allow the hydrolysis of the substrate by the enzyme. The intensity of the color generated is proportional to the amount of IgG specific antibody in the sample.

### Components

MATERIALS PROVIDED	96 Tests
1. Microwells coated with TG antigen	12x8x1
2. Sample Diluent: 1 bottle (ready to use)	22 ml
3. Enzyme conjugate: 1 bottle (ready to use)	12 ml
4. TMB Substrate: 1 bottle (ready to use)	12 ml
5. Calibrator: 1 Vial (ready to use)	1 ml

6. Positive Control: 1 vial (ready to use)	1 ml
7. Negative Control: 1 vial (ready to use)	1 ml
8. Stop Solution: 1 bottle (ready to use)	12 ml
9. Wash concentrate 20X: 1 bottle	25 ml

**Materials and Equipment Required but Not Provided**

1. Distilled or deionized water
2. Precision pipettes
3. Disposable pipette tips
4. ELISA reader capable of reading absorbance at 450nm
5. Absorbance paper or paper towel

**Disclaimer**

This product is for research use only and not intended for diagnostic procedures.

**Specimen Collection and Preparation**

1. Collect blood specimens and separate the serum.
2. Specimens may be refrigerated at 2–8°C for up to seven days or frozen for up to six months. Avoid repetitive freezing and thawing.

**Reagent Preparation**

1. Prepare 1X Wash buffer by adding Wash Concentrate (25 ml, 20X) to 475 ml of distilled or deionized water. Store at room temperature (18-26°C).

**Assay Procedure**

- Before proceeding with the assay, bring all reagents, serum references and controls to room temperature (18-26°C). Gently mix all reagents before use
  - The components in this kit are intended for use as an integral unit. The components of different lots should not be mixed
  - It is recommended that standards, control and serum samples be run in duplicate
  - Do not use sodium azide as preservative. Sodium azide inhibits HRP enzyme activities
1. Place the desired number of coated strips into the holder.
  2. Negative control, positive control, and calibrator are ready to use. Prepare 1:21 dilution of test samples, by adding 10 µl of the sample to 200 µl of sample diluent. Mix well.
  3. Dispense 100 µl of diluted sera, calibrator and controls into the appropriate wells. For the reagent blank, dispense 100 µl sample diluent in 1A well position. Tap the holder to remove air bubbles from the liquid and mix well. Incubate for 20 minutes at room temperature.
  4. Remove liquid from all wells. Wash wells three times with 300 µl of 1X wash buffer. Blot on absorbance paper or paper towel.
  5. Dispense 100 µl of enzyme conjugate to each well and incubate for 20 minutes at room temperature.
  6. Remove enzyme conjugate from all wells. Wash wells three times with 300 µl of 1X wash buffer. Blot on absorbance paper or paper towel
  7. Dispense 100 µl of TMB substrate and incubate for 10 minutes at room temperature.
  8. Add 100 µl of stop solution.

9. Read O.D. at 450 nm using ELISA reader within 15 min. A dual wavelength is recommended with reference filter of 600-650 nm.

### Calculation of Results

1. Check Calibrator Factor (CF) value on the calibrator bottle. This value might vary from lot to lot. Make sure you check the value on every kit.
2. Calculate the cut-off value: Calibrator OD x Calibrator Factor (CF).
3. Calculate the Ab (Antibody) Index of each determination by dividing the O.D. value of each sample by cut-off value.

### Example of a Standard Curve

Calibrator mean OD = 0.8

Calibrator Factor (CF) = 0.5

Cut-off Value =  $0.8 \times 0.5 = 0.400$

Positive control O.D. = 1.2

Ab Index =  $1.2 / 0.4 = 3$

Patient sample O.D. = 1.6

Ab Index =  $1.6 / 0.4 = 4.0$

### Quality Control

The test run may be considered valid provided the following criteria are met:

1. If the O.D. of the Calibrator should be greater than 0.250.
2. The Ab index for Negative control should be less than 0.9.
3. The Ab index for Positive control should be greater than 1.2.

### Interpretation

The following is intended as a guide to interpretation of TG antibody test results; each laboratory is encouraged to establish its own criteria for test interpretation based on sample populations encountered.

#### • Antibody Index Interpretation

- $<0.9$  No detectable antibody to TG by ELISA
- $0.9-1.1$  Borderline positive. Follow-up testing is recommended if clinically indicated.
- $>1.1$  Detectable antibody to TG by ELISA

#### • Converting of Ab Index to IU/mL

As an option, TG Ab index may be converted to IU/mL by multiplying Ab index value by 100. International units may then be interpreted as follows:

- $<90$  IU/mL: Negative
- $90-110$  IU/mL: Borderline positive
- $>110$  IU/mL: Positive

### References

1. Fan JL; Patibandla SA; Kimura S; Rao TN; Desai RK; Seetharamaiah GS; Kurosky A; Prabhakar. BS Purification and characterization of a recombinant human thyroid peroxidase expressed in insect cells. J Autoimmun 1996; 9(4):529-36.
2. Feldt-Rasmussen U. Analytical and clinical performance goals for testing autoantibodies to thyroperoxidase, thyroglobulin, and thyrotropin receptor. Clin Chem 1996;42:160-3.

3. Franke WG; Schimming C; Wunderlich G. Can thyroid peroxidase be used as a complementary tumor marker besides thyroglobulin? Preliminary experience with determination of TPO in differentiated thyroid carcinomas. *Anticancer Res* 1997;17(4B):2999-3002.
4. Haapala AM; Hyöty H; Parkkonen P; Mustonen J; Soppi E. Antibody reactivity against thyroid peroxidase and myeloperoxidase in autoimmune thyroiditis and systemic vasculitis. *Scand J Immunol* 1997; 46(1):78-85.
5. Mariotti S, Caturegli P, Piccolo P, Barbesino G, Pinchera A. Antithyroid peroxidase autoantibodies in thyroid diseases. *J Clin Endocrinol Metab* 1990;71:661-9.
6. Massart C, Guilhem I, Gibassier J, Allannire H, Nicol M. Comparison of thyroperoxidase and microsomal antibody assays in sera from patients with Graves disease. *Clin Chem* 1991;37:1777-80.
7. Nakamura H; Genma R; Mikami T; Kitahara A; Natsume H; Andoh S; Nagasawa S; Nishiyama K; Chida K; Sato A; Yoshimi T. High incidence of positive autoantibodies against thyroid peroxidase and thyroglobulin in patients with sarcoidosis. *Clin Endocrinol (Oxf)* 1997; 46(4):467-72.
8. Roti E, Gardini E, Minelli R, Bianconi L, Braverman LE. Prevalence of anti-thyroid peroxidase antibodies in serum in the elderly: comparison with other tests for anti-thyroid antibodies. *Clin Chem* 1992;38:88-92.

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