

Human CD36 ELISA Kit

Catalog No. EA200060

Principle of the Assay

CD36 (cluster of differentiation 36), also known as fatty acid translocase (FAT), platelet glycoprotein 4 (GPIV), scavenger receptor class B member 3 (SR-B3), and glycoproteins 88 (GP88), is an integral membrane protein found on the surface of many cell types in vertebrate animals. CD36 is a multiligand pattern recognition receptor that interacts with a large number of structurally dissimilar ligands such as collagen, thrombospondin, oxidized low density lipoprotein, native lipoproteins, oxidized phospholipids, and long-chain fatty acids. Upon ligand binding, CD36 transduces signals that mediate a wide range of pro-inflammatory cellular responses. CD36 may be involved in glucose intolerance, hypertension, atherosclerosis. arterial diabetes, cardiomyopathy, Alzheimer's disease and various cancers.

This sandwich ELISA is used to measure human CD36 in serum, plasma, and cell culture supernates. Microtitration wells coated with anti-human CD36 capture antibody are exposed to test specimens. The CD36 antigen in the specimen is specifically captured onto the immobilized antibody during specimen incubation. The captured CD36 antigen is then reacted with a biotinylated human CD36 detection antibody. Subsequently, Streptavidin-HRP conjugate is then added. After wash, specifically bound enzyme conjugate is detected by reaction with the Substrate Solution, tetramethylbenzidine (TMB). The assay is measured spectrophotometrically to indicate the level of CD36 present in a sample.

Materials Supplied

Description	Quantity	
CD36 Monoclonal Antibody Coated 96-well Plate in foil pouch with desiccant	1	
Recombinant Human CD36 Protein (5000ng/mL)0.1 mL		
Biotinylated CD36 Monoclonal Antibody (100x)	120 μL	
Streptavidin -HRP Conjugate (100x)	120 μL	
Standard Diluent	10 mL	
Sample Diluent	30 mL	
Assay Buffer	30 mL	
Substrate Solution (TMB)	12 mL	
Stop Solution (1N HCl)	12 mL	
Wash Buffer (20x)	60 mL	
Plate Sealer	3	

Additional Materials not Supplied

 Horizontal orbital plate shaker capable of maintaining a speed of 450±50 rpm.

- Disposable tip micropipettes to deliver volumes of 5μL, 10 μL, 25 μL, 100 μL and 200 μL (multichannel pipette preferred for dispensing reagents into microtiter plates).
- 3. Distilled or deionized water.
- Clean, disposable plastic/glass test tubes, approximate capacities 5mL and 10mL.
- Range of standard, clean volumetric laboratory glassware consisting of, at least, 15 mL and 100 mL beakers, 1 L graduated cylinder, 1 mL, 5 mL, and 10 mL pipettes.
- 6. Absorbent paper towels.
- 7. Automatic microplate washer or laboratory wash bottle.
- 8. Microplate reader with 450nm filter.
- Latex gloves, safety glasses and other appropriate protective garments.
- 10. Biohazard waste containers.
- 11. Safety pipetting devices for 1 mL or larger pipettes.
- 12. Timer.

Storage and Stability

Upon receipt, store the kit at 2-8°C. The kit should not be used beyond the expiration date. Once opened, the unused microplate strips should be returned to their original foil pouch along with the desiccant. The diluted Wash Buffer should not be stored for longer than 3 weeks at 2-8°C. It is recommended that Wash buffer be freshly diluted before each assay. If the diluted Wash buffer becomes visibly cloudy during the 3 weeks, discard it. (Note: Concentrated Wash Buffer, when stored at 2-8°C, normally may develop crystalline precipitates, which can be re-dissolved at 37°C.)

Indications of Deterioration

The human CD36 Assay kit may be considered to have deteriorated if:

- 1. Reagents are visibly cloudy.
- 2. The Substrate Solution turns blue. This is likely to be caused by chemical contamination of the Substrate Solution.

Precautions

- 1. The reagents supplied in this kit are for Research use only.
- 2. All blood products should be treated as potentially infectious.
- 3. Disposal or decontamination of fluid in the waste reservoir should be in accordance with guidelines described in the Department of Labor, Occupational Safety and Health Administration, occupational exposure to blood-borne pathogens; final rule (29 CFR 1910,1030) FEDERAL REGISTER, pp. 64176-84177,12/6/91.
- 4. The Substrate Solution and Stop Solution in this kit can irritate the skin and cause eye damage. Handle them with care and wear protective gloves, clothing and eye/face protection. Wash hands thoroughly after handling. Immediately flush the affected area with plenty of water in case of contact with skin or eyes. Obtain medical attention if necessary.

Technical Suggestions

- 1. This kit should be used in strict accordance with the instructions in the Package Insert.
- Do not use the kit after the expiration date printed on the outer carton label.



- 3. Do not cross contaminate reagents.
- Some reagents in the CD36 ELISA kit are optimized for each kit lot. Do not exchange reagents from kits with different lot numbers.
- 5. To ensure accurate results and avoid crosscontamination, use proper adhesive plate sealers during incubation steps, and change pipette tips when adding each standard and sample. Multi-channel pipettes are recommended for large assays. Always use fresh pipette tips when drawing from stock reagent bottles.
- Warm up the foil bag to room temperature before opening.
- All reagents should be added to the plate in the same order.
- 8. If the Stop Solution does not mix thoroughly with the Substrate Solution, the color in the wells may appear green after adding stop solution. Gently tap the plate or pipette up and down to mix until the color in the wells change to yellow (avoid bubbles during this step).
- Reagents should be dispensed with the tip of the micropipettes touching the side of the well at a point about mid-section. For automatic processors, follow manufacturer's recommendations.
- It is recommended that all pipetting devices (manual or automatic), and thermometers are regularly calibrated according to the manufacturer's instructions.

Sample Collection and Storage

The Human CD36 ELISA is intended for use with serum, plasma, and other biological fluids. The specimen should be tested as soon as possible. However, if the specimen needs storage, the specimens should be stored frozen at -20°C or below. Do not use self-defrosting freezers. Specimens that have been frozen and thawed should be thoroughly mixed before testing.

Cell Culture Supernatants - Remove particulates by centrifugation and assay immediately or aliquot and store samples at \leq -20 °C. Avoid repeated freeze-thaw cycles.

Serum - Use a serum separator tube (SST) and allow samples to clot for 30 minutes at room temperature before centrifugation for 15 minutes at $1000 \times g$. Remove serum and assay immediately or aliquot and store samples at \le -20 °C. Avoid repeated freeze-thaw cycles.

Plasma - Collect plasma using EDTA or heparin as an anticoagulant. Centrifuge for 15 minutes at $1000 \times g$ within 30 minutes of collection. Assay immediately or aliquot and store samples at \leq -20 °C. Avoid repeated freeze-thaw cycles.

Rinse Cycle

Aspirate each well and wash. Wash by filling each well with Wash Buffer ($300\mu L$) using a squirt bottle, manifold dispenser, or automatic plate washer. Complete removal of liquid at each step is essential to good performance. After the last wash, invert the plate and blot it against clean paper towels.

Preparation for the Assay

1. Standard preparation: Prepare protein standard by diluting $10\mu L$ of standard stock into 490 μL (1:50 dilution) of Standard Diluent. This will give a final concentration of 100 ng/mL as shown in Table 1. Make 2.5x serial dilution of Standard 1 using Standard Diluent. to generate a standard concentration range of 0.41 to 100 ng/mL.

- 2. Sample preparation: CD36 concentration must be estimated prior to performing the full experiment by testing a serially diluted representative sample using **Sample Diluent**. Select an optimal dilution level such that the final target protein concentration falls near the middle of the assay linear dynamic range.
- 3. Detection antibody preparation: dilute the concentrated biotin conjugated detection antibody 1:100 using Assay Buffer.
- 4. *SA-HRP preparation*: dilute the concentrated streptavidin HRP conjugate 1:100 using **Assay Buffer**.
- 5. Wash buffer: Prepare working-strength Wash buffer by diluting 1 part concentrate with 19 parts of distilled or deionized water. If a kit is likely to be utilized over a period in excess of 4 weeks, then it is recommended that only enough stock concentrate be diluted sufficient for immediate needs.

Table 1: Human CD36 Standard Curve Generation

Standard Number	Concentration of CD36 (ng/mL)	CD36 Standard (µL)	Standard Diluent (µL)
1	100	10	490
2	40	160 of #1	240
3	16	160 of #2	240
4	6.40	160 of #3	240
5	2.56	160 of #4	240
6	1.02	160 of #5	240
7	0.41	160 of #6	240
8	0		240

Assay Procedure

Note: All standards, controls and samples should be tested in duplicate.

- 1. Allow all reagents to reach room temperature (18-25°C).
- 2. Select sufficient microtitration well strips to accommodate all test specimens, controls and reagent blank. Fit the strips into the holding frame.
- 3. Dispense 100 $\,\mu L$ of each standard and sample into appropriate wells.
- 4. Incubate for 2 hours at room temperature with moderate shaking (450±50rpm) on a horizontal orbital plate shaker.
- 5. Wash the microtitration plate 3 times as described in the Rinse Cycle section.
- 6. Add 100 μ L of working concentration detection antibody into each well and incubate for 1.5 hour at room temperature with moderate shaking (450±50rpm) on a horizontal orbital plate shaker.
- 7. Wash the microtitration plate 3 times as described in the Rinse Cycle section.
- 8. Add 100 μ L of working concentration Streptavidin HRP conjugate into each well and incubate for 25 minutes at room



temperature with moderate shaking (450±50rpm) on a horizontal orbital plate shaker.

- 11. Wash the microtitration plate 5 times as described in the Rinse Cycle section.
- 12. Dispense 100 µL Substrate Solution into each well. A multichannel pipette should be used for best results. Leave at room temperature (18-25°C) and protected from direct sunlight for 20-25 minutes.
- 13. Stop the reaction by adding 100 μL of Stop Solution to each well. The blue solution should change to a uniform yellow color. Ensure that the undersides of the wells are dry and that there are no air bubbles in the well contents.
- 14. Read the absorbance values at 450 nm using a microplate reader. If wavelength correction is available, set to 540 nm or 570 nm.

Calculation of Results

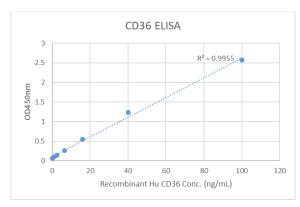
Average the duplicate readings for each standard and sample. A 4-parameter logistic (4-PL) or a linear regression model providing a point-to-point curve fitting provides acceptable results. Create a standard curve by reducing the data using computer software capable of generating a fourparameter logistic (4-PL) or a linear regression curve-fit. As an alternative, construct a standard curve by plotting the mean absorbance for each standard on the y-axis against the concentration on the x-axis and draw a best fit curve through the points on the graph. Do not force the line to be linear. The concentration of the samples can be found directly from the standard curve.

Table 2. Example Data at 450nm.

Standards	450 nm absorbance
Standard 1 (100ng/mL)	2.572
Standard 2 (40ng/mL)	1.235
Standard 3 (16ng/mL)	0.556
Standard 4 (6.4ng/mL)	0.267
Standard 5 (2.56ng/mL)	0.147
Standard 6 (1.02ng/mL)	0.102
Standard 7 (0.41ng/mL)	0.082
Standard 8 (0 ng/mL)	0.062

Typical Human CD36 ELISA Kit Standard Curve

This standard curve was generated at OriGene for demonstration purpose only.



Note: This standard curve is only an example and should not be used to generate any results.

Assay Sensitivity: 0.263ng/mL

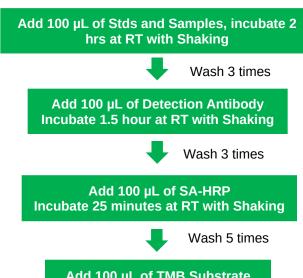
Limitations of Use

- 1. This kit is for research use only, not for use in diagnostic procedures.
- 2. The CD36 value measured using OriGene CD36 ELISA kit may not be interchangeable with that obtained from other assay kits.
- 3. The assay cannot be used to quantitate samples with values higher than the highest standard without further dilution of the samples.

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Assay Flowchart



Add 100 µL of TMB Substrate Incubate 20-25 minutes at RT



Add 100 µL of Stop Solution



Read OD450nm in Plate Reader