Mouse IgG RapidTiter Strips

Catalog No.
AR100097 (25 Strips)
AR100097L (100 Strips)

Principle of the Assay

Mouse IgG RapidTiter Strips are ready-to-use Dip & Read test strips, designed for rapid detection and semi quantitative measurement of mouse IgG, and mouse IgG Fc fused proteins. Just immerse the strip in your sample and interpret the result in less than five minutes—no specialized equipment or instruments required. The strip can be applied directly to unpurified samples in cell lysis buffer or cell culture medium (with or without FBS). It is also compatible with samples in common buffers such as PBS, Tris, or HEPES etc.

The Mouse IgG RapidTiter Strip detects can detect all mouse IgG subtypes including IgG1, IgG2a, IgG2b and IgG3, with a sensitivity of 0.1 µg/mL. The quantitative range extends from the detection limit up to 5 mg/mL. Unlike traditional sandwich lateral flow assays, this test is not affected by the hook effect and does not produce false-negative results in samples with high analyte concentrations. By comparing the line pattern with the reference images provided in the protocol, the sample concentration can be semi-quantitatively determined by simple visual inspection with the naked eye. Mouse IgG RapidTiter Strips are intended for research use only.

Package Contents

Cat #	RapidTiter Strips	Reference Card
AR100097	25 Strips	1
AR100097L	4 x 25 Strips	1

Storage and Stability

Store all strip components at room temperature. Strips should not be used beyond the expiration date. Leave RapidTiter Strips in its canister until just before use. Keep dry. Avoid prolong exposure to humidity.

Precautions

1. The test strip contains a total of three lines: a control line (C), test line T1, and test line T2. The control line (C) is the uppermost line, located approximately 42 mm from the bottom of the strip. Test line T1 is the middle line, positioned about 38 mm from the strip bottom, and test line T2 is the bottom line, located approximately 34 mm from the strip bottom.
2. The line patterns shown in the protocol are read for 5 minutes. Reading the results at longer time points may increase assay sensitivity. The line pattern remains stable for at least 3 hours, although the line color may gradually fade over time.
3. Results for each mouse IgG subtype are provided in this manual. Although the sensitivity is similar among subtypes, the line patterns may differ at certain concentrations. Users should therefore compare results with the reference images corresponding to the same IgG isotype.
4. This product can also be used to detect mouse IgG Fc–fused proteins. The concentration of mouse IgG Fc–fused proteins should be calculated according to the molecular weight of the fusion protein.

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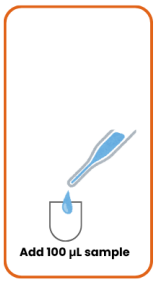
5. For more accurate measurements, standard samples with known concentrations of the same protein in the same matrix can be used to first generate reference line patterns. The sample results can then be compared with these reference patterns.

Test Procedure

Read all instructions carefully before performing the test. Failure to follow the instructions may result in inaccurate test results.

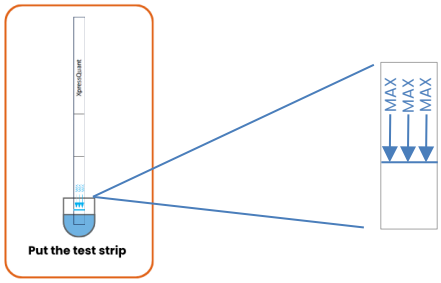
1. Add 100 µL sample to a well (e.g. a well of a 96-well plate) or a vial.

   Note: Dilute the sample with a buffer (e.g. PBS) if desired.



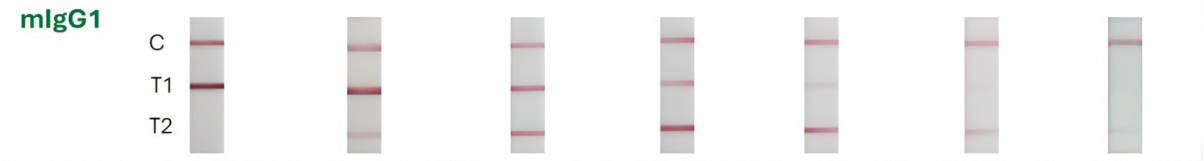
2. Put the test strip into the vial, with the arrow label side immersed in the sample.

   Note: Do not immerse above the MAX line



3. Wait 5 minutes until the control line is clear.

4. Read the result by comparing the images below



Concentration	0	0.1 µg/mL	1 µg/mL	10 µg/mL	100 µg/mL	1 mg/mL	≥ 5 mg/mL
Line pattern	Two clear lines C and T1, T1 > C	Two clear lines C, T1 and a weak T2. T1 ≥ C > T2	Three clear lines C, T1 and T2. T1 ≥ T2 ≈ C	Three clear lines C, T1 and T2. T2 > C ≈ T1	Two clear lines C, T2 and a weak T1. T2 ≈ C > T1	Two clear lines C, T2 and a faint T1. C > T2 >> T1	A clear C line and faint T2, T1 lines. C >> T2 > T1

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mIgG2a

Concentration	0	0.1 μg/mL	1 μg/mL	10 μg/mL	100 μg/mL	1 mg/mL	≥ 5 mg/mL
Line pattern	Two clear lines C and T1, T1 > C	Two clear lines C, T1 and a faint T2. T1 > C >> T2	Three clear lines C, T1 and T2. T1 ≥ C > T2	Three clear lines C, T1 and T2. C ≈ T2 ≈ T1	Two clear lines C, T2 and a faint line T1 C ≈ T2 >> T1	A clear C line and a weak faint line. T1 is invisible. C >> T2	A clear C and faint or invisible T2 , C >> T2

mIgG2b

Concentration	0	0.1 μg/mL	1 μg/mL	10 μg/mL	100 μg/mL	1 mg/mL	≥ 5 mg/mL
Line pattern	Two clear lines C and T1, T1 > C	Two clear lines C, T1 and a faint T2. T1 > C >> T2	Three clear lines C, T1 and T2. T1 ≥ C > T2	Three clear lines C, T1 and T2. C > T1 ≈ T2	Two clear lines C, T2 and a faint line T1 C ≈ T2 >> T1	A clear C line and a faint T2 line. T1 is invisible. C >> T2	Only one line C

mIgG2c

Concentration	0	0.1 μg/mL	1 μg/mL	10 μg/mL	100 μg/mL	1 mg/mL	≥ 5 mg/mL
Line pattern	Two clear lines C and T1, T1 > C	Two clear lines C, T1 and a faint T2. T1 > C >> T2	Three clear lines C, T1 and T2. T1 ≥ C > T2	Three clear lines C, T1 and T2. C ≥ T2 ≈ T1	Two clear lines C, T2 and a faint line T1 C > T2 >> T1	A clear C line and a faint T2 line. T1 is invisible. C >> T2	Only one line C

mIgG3

Concentration	0	0.1 μg/mL	1 μg/mL	10 μg/mL	100 μg/mL	1 mg/mL	≥ 5 mg/mL
Line pattern	Two clear lines C and T1, T1 > C	Two clear lines C, T1 and a faint T2. T1 > C >> T2	Three clear lines C, T1 and T2. T1 ≥ C > T2	Three clear lines C, T1 and T2. C ≥ T2 ≈ T1	Two clear lines C, T2 and a weak line T1 C ≈ T2 > T1	Two clear lines C, T2 and a faint T1. T2 ≥ C >> T1	Two clear lines C, T2. T1 is invisible. T2 ≥ C

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