

## In vitro digestion of DNA with Cas9 nuclease and sgRNA

### Overview:

This protocol describes how to digest double-stranded DNA *in vitro* using OriGene Cas9 protein (TP790148) and a synthetic guide RNA (sgRNA).

### Required Materials:

- Cas9 Nuclease, *S. pyogenes* (Cat. No. TP790148)
- 10X Cas9 Nuclease Reaction Buffer (20mM HEPES, 100mM NaCl, 5mM MgCl<sub>2</sub>, 0.1mM EDTA, pH 6.5 at 25°C, pH is critical for Cas9 activity, make sure buffer has the right pH value).
- Nuclease-free water
- sgRNA containing the targeting sequence
  - ✓ Synthetic pure sgRNA can be conveniently ordered from OriGene sgRNA service. <http://www.origene.com/CRISPR-CAS9/>
  - ✓ Alternatively, sgRNAs can be generated by *in vitro* transcription Kit using linearized plasmid, PCR products, or oligonucleotides as templates
  - ✓ sgRNAs must contain sequence complementary to the target DNA (guide sequence)
- DNA substrate
  - ✓ Substrate can be circular or linearized plasmid, PCR products, or synthesized oligonucleotides. Linearized DNA is preferred.
- Equipment and Reagents for DNA fragment analysis (agarose, DNA loading dye, Gel electrophoresis apparatus, etc.).

### Before Starting

- We strongly recommend wearing gloves and using nuclease-free tubes and reagents to avoid RNase contamination.
- Reconstitution of synthetic sgRNA
  - Spin down briefly before opening the tube.
  - Add 100μl nuclease-free water to make 10μM stock solution.
  - Dilute stock solution to make 1μM working solution.
- Prepare 200-400 ng linearized DNA as substrate.

## Procedure

1. Assemble the reaction in a nuclease free PCR tube in the following order:

Component	Volume (30ul final)
Nuclease-free water	21 $\mu$ l
10X Cas9 Nuclease Reaction Buffer	3 $\mu$ l
1 $\mu$ M sgRNA	1 $\mu$ l (~30 nM final)
1 $\mu$ M Cas9 Nuclease, <i>S. pyogenes</i> (TP790148)*	1 $\mu$ l (~30 nM final)
20 nM substrate DNA	4 $\mu$ l (~3 nM final)
Total reaction volume	30 $\mu$ l

\* 1  $\mu$ M equals 165 ng/ $\mu$ l.

2. Mix thoroughly and spin down briefly.
3. Incubate at 37°C for 1 hour, then heat at 65°C for 10 min to deactivate Cas9 nuclease. Hold the reactions at 4°C if you want to take a break.
4. Proceed with fragment analysis by gel electrophoresis.