

Suggested Transformation Protocol:

- 1. Thaw cells on wet ice after removing from -70°C
- Mix cells gently by lightly flicking the tube. Aliquot ~50-100µl of cells into chilled, 17 x 100mm polypropylene tube(s). Unused cells may be refrozen, but a small drop in efficiency may result. For optimal recovery, refreeze cells in a dry ice/ ethanol bath prior to -70°C storage.
- 3. Add DNA solution (≤5µl per 50µl cells) to cell suspension and gently swirl tube(s) for a few seconds to mix. If a control is desired, repeat this step with 2µl of the provided pUC19 in a separate tube.
- 4. Incubate on ice for 30 minutes.
- 5. Place tube(s) in 42°C water bath for ~30 to 45 seconds without shaking.
- 6. Place tube(s) again on ice for ~2 minutes.
- 7. Dilute transformation reaction(s) to 1ml by addition of 900-950µl SOC medium**.
- 8. Shake tube(s) ~200 rpm for 60 minutes at 37°C.
- 9. Plate by spreading 5-200µl of cell transformation mixture on LB agar plates containing appropriate antibiotic and incubate overnight at 37°C.

When performing the pUC19 control transformation, plate 5µl of the transformation mixture on a LB agar plate containing 100µg/ml ampicillin. To facilitate cell spreading, place a pool of SOC (100µl) onto surface of plate prior to addition of transformation mixture.

 ** SOC Medium: 2% Tryptone, 0.5% Yeast Extract, 0.4% glucose, 10mM NaCl, 2.5mM KCl, 10mM ${\rm MgCl}_2$ & 10mM ${\rm MgSO}_4$.

Transformation Efficiency Calculation for Control DNA

For example:

If 40 colonies were obtained after transforming 20pg of pUC19 and plating 5µl of the final 1ml transformation mixture, the calculated transformation efficiency would be: