

CD9 Exosome (EV) Capture Kit

Quick Start Guide

What's in the Package

- User Manual
- Magnetic beads
- 10X Wash buffer

Storage Requirement

- 2–8°C
- Do not freeze

Quick Protocol:

1. Sample Preparation

Plasma/Serum	Cell Culture Medium
<ul style="list-style-type: none">• 300–500 × g, 10 min → remove cells• 3,000 × g, 20–30 min → remove debris• 10,000 × g, 30 min → remove platelets• Collect pre-cleared sample	<ul style="list-style-type: none">• Collect conditioned medium• 300–500 × g, 10 min → remove cells• 3,000 × g, 30 min → remove debris/large vesicles

2. Bead Preparation

- a. Vortex beads for 30 seconds
- b. Aliquot 50 uL beads into low-binding tube and add 0.5 mL PBS+0.1% BSA
- c. Place on magnetic rack for >30 second, remove supernatant

Beads are ready for binding

3. Binding

- a. Add 200 uL of EV sample (pre-cleared plasma, serum, cell culture media, pre-enriched EV) to beads. **Tip:** for RNA workflows, add RNase inhibitor (final 0.5–1U/mL) at this step
- b. Incubate 2 hr at RT or overnight at 4°C with gentle mixing
- c. Place on magnetic rack for > 30 second, remove supernatant

CD9+ EVs are now bound to beads

4. Wash (Protein analysis: 2–3 washes; Nucleic acids: optional)

- a. Add 500 uL of 1X Wash Buffer to the bound beads
- b. Resuspend beads (vortex + quick spin)
- c. Place on magnetic rack for >30 seconds, remove wash buffer
- d. Repeat 1–2 times

Clean CD9+EVs bounds to beads are ready for downstream applications

Additional Notes

- 200 µL is a recommended starting sample volume. Because EV abundance can vary by sample type and source, users should optimize performance by titrating the bead-to-sample ratio through input bead amount and sample volume. Reaction volumes may be scaled as needed.
- Proper mixing is critical—ensure beads remain suspended. Use appropriate size of reaction tube and a mixer that tilts and rotates to ensure beads do not settle at the bottom of tube.

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