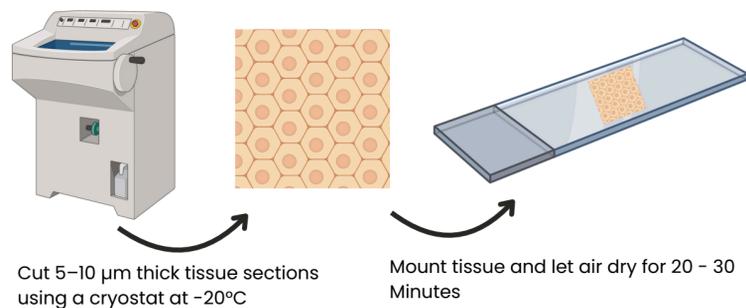


Protocol

CHROMOGENIC IMMUNOHISTOCHEMISTRY STAINING OF FROZEN TISSUE

1. Tissue Sectioning & Fixation

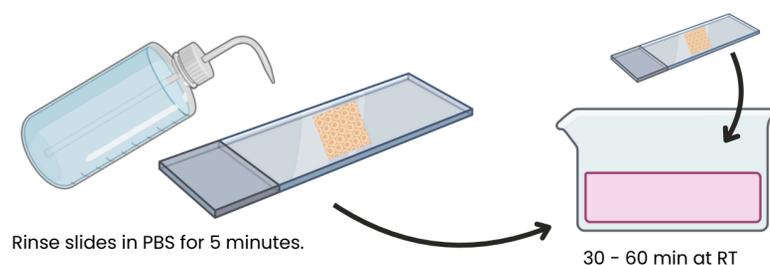


Fix sections:

Option A (acetone): Fix in cold acetone (-20°C) for 10 minutes.

Option B (PFA): Fix in 4% PFA for 10 minutes, then rinse in PBS. Air dry the slides again for 10 minutes.

2. Permeabilization & Blocking

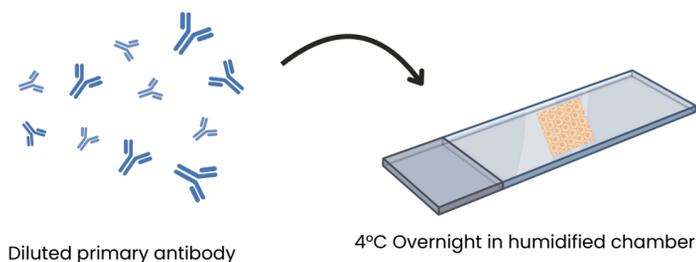


Rinse slides in PBS for 5 minutes.

(Optional) Incubate with 0.1–0.3% Triton X-100 in PBS for 10 minutes if intracellular targets are being stained.

Block with 5–10% normal serum (same species as the secondary antibody) or 1–5% BSA in PBS for 30–60 minutes at room temperature.

3. Primary Antibody Incubation



Dilute the primary antibody as recommended by the manufacturer.

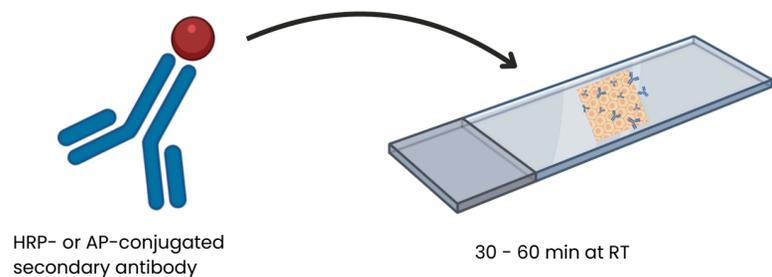
Apply to each section and incubate:

1–2 hours at room temperature or

Overnight at 4°C in a humidified chamber.

Rinse in PBS, 3 × 5 minutes.

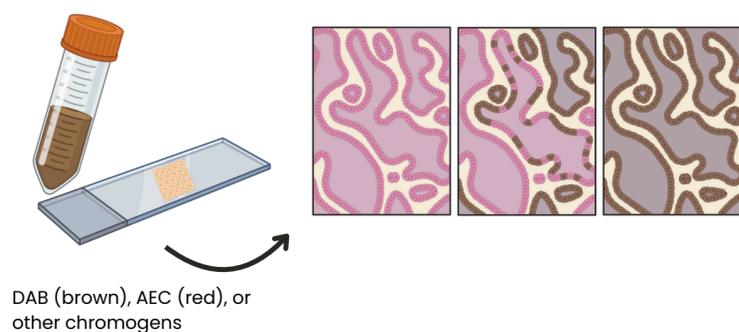
4. Secondary Antibody & Detection



Incubate sections with the enzyme-conjugated secondary antibody for 30–60 minutes at room temperature.

Rinse in PBS or TBS, 3 × 5 minutes.

5. Chromogenic Substrate Development



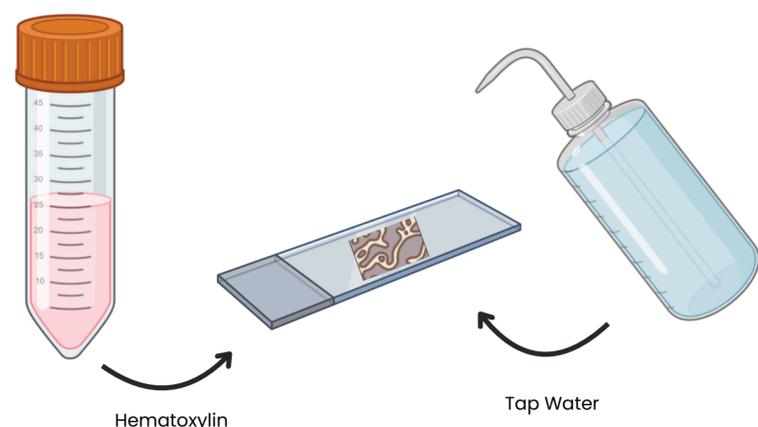
Prepare the chromogen fresh just before use.

Apply substrate to sections and monitor color development under a microscope (typically 5–15 minutes).

Stop the reaction by rinsing in distilled water.

⚠ Note: AEC is alcohol-soluble; avoid alcohol-based mounting.

6. Counterstaining & Mounting



Counterstain with hematoxylin for 30–60 seconds.

Rinse in tap water and blueing reagent if required.

Mount:

For DAB: Dehydrate, clear in xylene, and mount with a permanent medium.

For AEC: Mount with aqueous mounting medium (do not dehydrate).